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# Cytotoxic Effects of Dimorfolido-*N*-Trichloroacetylphosphorylamide and Dimorfolido-*N*-Benzoylphosphorylamide in Combination with C<sub>60</sub> Fullerene on Leukemic Cells and Docking Study of Their Interaction with DNA

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## Abstract

Dimorfolido-*N*-trichloroacetylphosphorylamide (HL1) and dimorfolido-*N*-benzoylphosphorylamide (HL2) as representatives of carbacylamidophosphates were synthesized and identified by the methods of IR, <sup>1</sup>H, and <sup>31</sup>P NMR spectroscopy. In vitro HL1 and HL2 at 1 mM concentration caused cell specific and time-dependent decrease of leukemic cell viability. Compounds caused the similar gradual decrease of Jurkat cells viability at 72 h (by 35%). HL1 had earlier and more profound toxic effect as compared to HL2 regardless on leukemic cell line. Viability of Molt-16 and CCRF-CEM cells under the action of HL1 was decreased at 24 h (by 32 and 45%, respectively) with no substantial further reducing up to 72 h. Toxic effect of HL2 was detected only at 72 h of incubation of Jurkat and Molt-16 cells (cell viability was decreased by 40 and 45%, respectively).

It was shown that C<sub>60</sub> fullerene enhanced the toxic effect of HL2 on leukemic cells. Viability of Jurkat and CCRF-CEM cells at combined action of C<sub>60</sub> fullerene and HL2 was decreased at 72 h (by 20 and 24%, respectively) in comparison with the effect of HL2 taken separately.

In silico study showed that HL1 and HL2 can interact with DNA and form complexes with DNA both separately and in combination with C<sub>60</sub> fullerene. More stable complexes are formed when DNA interacts with HL1 or C<sub>60</sub> + HL2 structure. Strong stacking interactions can be formed between HL2 and C<sub>60</sub> fullerene. Differences in the types of identified bonds and ways of binding can determine distinction in cytotoxic effects of studied compounds.

**Keywords:** Dimorfolido-*N*-trichloroacetylphosphorylamide, Dimorfolido-*N*-benzoylphosphorylamide, C<sub>60</sub> fullerene, Leukemic cells, DNA, Computer modeling

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## Background

Nowadays, it is important to create new biocompatible nanomaterials that exhibit pharmacological properties, antitumor activity, and modulate the biological effects of chemotherapeutic drugs.

Carbacylamidophosphates as the structural analog of  $\beta$ -diketones, that are promising class of biologically active compounds with antimitotic and antiproliferative activities, can be used as antitumor agents [1–3].

In previous study with the use of *in silico* analysis, we have shown that dimethyl-*N*-(benzoyl)amidophosphate as representative of carbacylamidophosphates possesses an antitumor activity and interacts with DNA [4, 5].

*In vitro* toxic effects (decrease of cell viability and increase of ROS production) of 2.5 mM dimethyl-*N*-(benzoyl)amidophosphate on leukemic L1210 cells were demonstrated. It was shown that toxic effects of dimethyl-*N*-(benzoyl)amidophosphate on leukemic cells could be facilitated by C<sub>60</sub> fullerene [5].

C<sub>60</sub> fullerene is a chemically stable carbon nanostructure, able to interact with biomolecules and penetrate through plasma membrane inside the cell [6–8] and, thus, it can be used in biomedical applications [9–13]. C<sub>60</sub> fullerene can form complexes with chemotherapeutic drugs such as doxorubicin, cisplatin, and paclitaxel, reducing their cytotoxic effect and enhancing the therapeutic effect [14–18].

Two carbacylamidophosphate derivatives with different substituents were used in this study: dimorfolido-*N*-trichloroacetylphosphorylamide (HL1) contained CCl<sub>3</sub> group (Fig. 1a) and dimorfolido-*N*-benzoylphosphorylamide (HL2)—benzene ring (Fig. 1b) at carbamide group.

The aim was to study the leukemic cell viability under the action of HL1 and HL2 separately and in combination with C<sub>60</sub> fullerene and to estimate their interaction with DNA *in silico*.

## Methods

### Chemicals

MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide], DMSO (Sigma-Aldrich Co, Ltd, USA).

### Characterization of Chemical Compounds

HL1 and HL2 were synthesized at the Department of Inorganic Chemistry, Chemical Faculty of Taras Shevchenko National University of Kyiv according to methods described earlier [19]. The composition of the synthesized compounds was confirmed by NMR and IR spectroscopy and measuring the melting points. The purity of above compounds was  $\geq 98\%$ .

IR measurements were performed on a UR-10 and a Perkin-Elmer Spectrum BX spectrometer on samples as KBr pellets. <sup>1</sup>H and <sup>31</sup>P NMR spectra in DMSO-d<sub>6</sub> and methanol-d<sub>4</sub> solutions were obtained on a Varian 400 NMR spectrometer at room temperature. Chemical shifts are referenced to SiMe<sub>4</sub> as interior standard for <sup>1</sup>H NMR and H<sub>3</sub>PO<sub>4</sub> as exterior standard for <sup>31</sup>P NMR.

HL1: m. p. 205 °C; IR (KBr, cm<sup>-1</sup>):  $\nu = 3013$  (NH), 1728 (CO), 1201 (PO); <sup>1</sup>H NMR (DMSO-d<sub>6</sub>): 3.61 (m, 8H, CH<sub>2</sub> $\alpha$ ), 3.17 (m, 8H, CH<sub>2</sub> $\beta$ ).

HL2: m. p. 178 °C; (KBr, cm<sup>-1</sup>):  $\nu = 3100$  (NH), 1685 (CO), 1200 (PO); <sup>1</sup>H NMR (methanol-d<sub>4</sub>): 3.36 (m, 8H, CH<sub>2</sub> $\alpha$ ), 3.24 (m, 8H, CH<sub>2</sub> $\beta$ ), 7.49, 7.56, 7.90 (m, C<sub>6</sub>H<sub>5</sub>) (8:8:5:1).

The synthesized compounds were dissolved in 10% DMSO to a final concentration of 0.05 M.

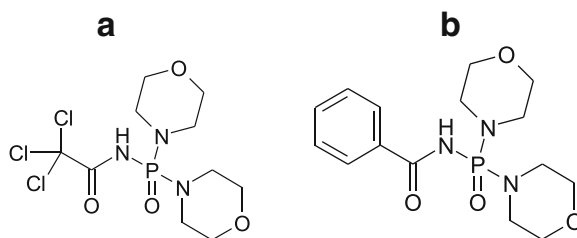
### Characterization of C<sub>60</sub> Fullerene Aqueous Solution

A highly stable aqueous colloid solution of C<sub>60</sub> fullerene (10<sup>-4</sup> M, purity >99.5%, nanoparticle average size up to 50 nm) was synthesized in Technical University of Ilmenau (Germany) as described in [20, 21].

### In Vitro Study

#### Cell Culture

The experiments were done on different human acute T cell leukemic cell lines. Cell lines were purchased from the Leibniz Institute DSMZ-German Collection of Microorganisms and Cell Cultures: Jurkat (ACC 282), CCRF-CM (ACC 240), and Molt-16 (ACC 29). The cells were cultured in RPMI 1640 medium supplemented with 10% FBS, 1% penicillin/streptomycin, and 2 mM glutamine using 25-cm<sup>2</sup> flasks at 37 °C with 5% CO<sub>2</sub> in humidified incubator.



**Fig. 1** Carbacylamidophosphates. **a** Dimorfolido-*N*-trichloroacetylphosphorylamide (HL1). **b** Dimorfolido-*N*-benzoylphosphorylamide (HL2)

Cells in RPMI 1640 medium were preincubated with  $C_{60}$  fullerene ( $10^{-5}$  M) during 1 h. After that, HL1 or HL2 was added to above medium and incubated for 24, 48, and 72 h.

Cell survival without addition of chemical compounds and  $C_{60}$  fullerene was received as 100% (control sample contained 0.05 M DMSO).

#### Cell Viability (MTT) Assay

Cell viability was assessed by the MTT reduction assay [22]. At indicated time points of incubation, 200  $\mu$ l aliquots ( $1 \times 10^5$  cells) was placed into the 96-well microplates, 20  $\mu$ l of MTT solution (4 mg/ml) was added to each well, and the plates were incubated for another 2 h. The culture medium was then replaced with 100  $\mu$ l of DMSO; diformazan formation was determined by measuring absorption at 570 nm with a microplate reader Tecan Infinite M200 Pro (Switzerland).

#### In Silico Study

The double-helix DNA molecule was used as a template from PDB (Protein Data Bank) base. The interactions of DNA molecule with HL1 or HL2 separately and in combination with  $C_{60}$  fullerene have been studied. We took into consideration the following structures of DNA molecule: 2MIW (CCATCGCTACC—intercalation of compound into a small groove of DNA helix), 1XRW (CCTCGTCC—intercalation of compound into a small groove of DNA helix), and 2M2C (GCGCATGC-TACGCG—binding of compound with large and small grooves of DNA helix). We applied the algorithm of systematical docking (SDOCK+), built-in the QXP package (this method demonstrates all possible conformations of the studied structures with the minimal value of root mean square deviation (Rmsd)) [23]. To each compound (HL1 or HL2) in combination with the  $C_{60}$  fullerene, we generated 300 potentially possible complexes with DNA, the 10 best of which were selected for the next stage, using a scoring function, built-in the QXP package [24].

The interactions of the DNA molecule with HL1 or HL2 separately and in combination with the  $C_{60}$  fullerene were characterized by the following parameters: (1) the number of hydrogen bonds, (2) the area of contacting surfaces of DNA and corresponding structure, (3) the distance between the DNA and docked structure, and (4) the total energy of the binding structure.

To assess the stability of the complexes of chemical compounds with  $C_{60}$  fullerene, we conducted the short-molecular dynamics (MD, 25 ps) using a Nosé-Poincaré-Anderson algorithm (NPA) [25, 26]. The calculations were performed on the following parameters: temperature (in K)—300; pressure (in kPa)—100;

binding involving the hydrogen atom or ligand was limited by the algorithm [26].

#### Statistical Analysis

The data were represented as  $M \pm SD$  of more than five independent experiments. Mean ( $M$ ) and standard deviation ( $SD$ ) were calculated for each group. Statistical analysis was performed using two-way ANOVA followed by post Bonferroni's tests. A value of  $p < 0.05$  was considered statistically significant. Data processing and plotting were performed by IBM PC using specialized applications GraphPad Prism 7 (GraphPad Software Inc., USA).

#### Results and Discussion

In the structures of HL1 and HL2, the carbonyl and phosphoryl groups are in *anti*-positions to each other as in most carbacylamidophosphates. In the structure of HL1, the bond lengths C–O (1.202(4) Å) and C–N (1.346(4) Å) are influenced by the substituent nature near the carbonyl group and have typical values for trichloroacetamide derivatives. The carbon atom of  $CCl_3$  group has a tetrahedral environment (Fig. 1a). For HL2, the planar benzene ring is rotated relative to the plane of the carbamide group on angle  $22.34(27)^\circ$ , which does not exclude the possibility of  $\pi$ -interaction between the benzene ring and the (O)CNP(O) fragment (Fig. 1b). It should be noted a close contact between the electrophilic phosphorus and nucleophilic oxygen atoms of the carbonylic group for both ligands. The distance between above-mentioned atoms 3.02 Å for HL1 and 3.122(3) Å for HL2 is slightly below than the sum of the Van der Waals radii of phosphorus and oxygen atoms (3.3 Å) that can be considered as an evidence of estimate covalent contribution in interatomic interaction [27, 28].

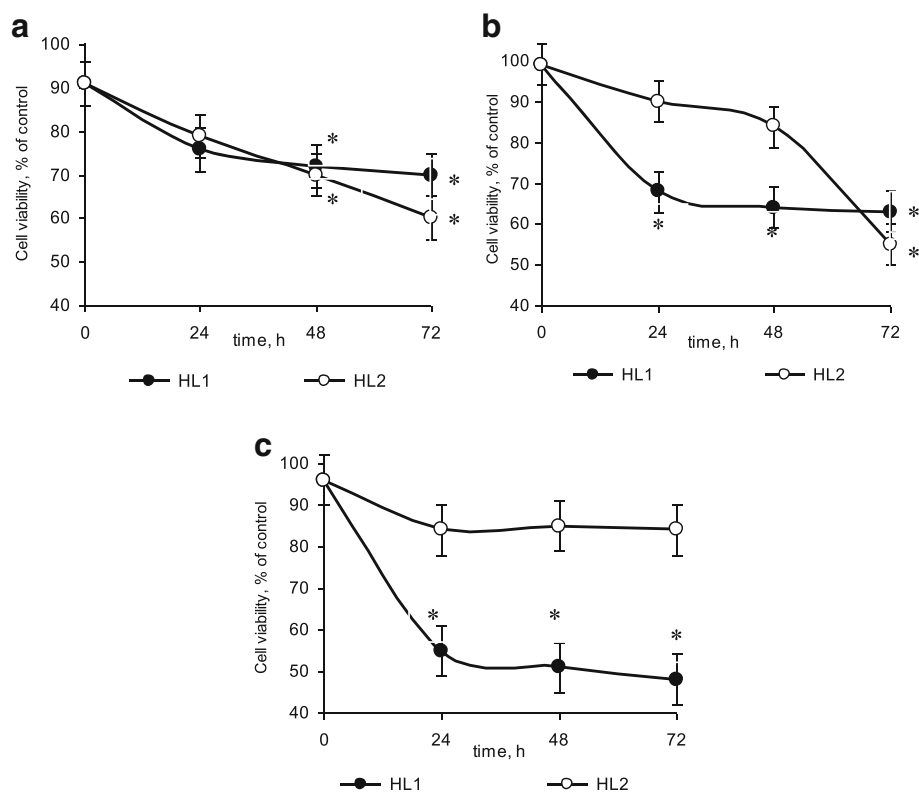
More electronegative character of  $CCl_3$  group in comparison with  $C_6H_5$  (phenyl) group reflects in the shortening of intramolecular hydrogen bonds N...O and O...H type: for HL1—1.81(1) Å and 2.73(1) Å, and for HL2—2.103(3) Å and 2.946(3) Å, respectively.

HL1 and HL2 at 1 mM concentration were screened for their toxicity against human T cell leukemic cells Jurkat, CCRF-CEM, and Molt-16 using MTT assay.

It was shown that HL1 and HL2 decrease the viability of leukemic cells. The observed toxic effect was cell specific and time dependent (Fig. 2).

HL1 and HL2 caused the similar gradual decrease of Jurkat cells viability by about 35% at 72 h (Fig. 2a).

The dynamics of Molt-16 and CCRF-CEM cell death under the action of HL1 was similar: cell viability was decreased at 24 h with no substantial further reducing up to 72 h. But the value of toxic effect was higher on



**Fig. 2** Viability of leukemic cells in the presence of 1 mM HL1 or HL2.  $M \pm m$ ,  $n = 8$ ;  $*p < 0.05$  compared to control cells. **a** Jurkat cells. **b** Molt-16 cells. **c** CCRF-CEM cells

CCRF-CEM cells (52% at 72 h) as compared with Molt-16 cells (37% at 72 h) (Fig. 2b, c).

Toxic effect of HL2 against Molt-16 cells was less pronounced as compared with HL1. Molt-16 cells viability was decreased (by 45%) only in 72 h, while no substantial decrease of CCRF-CEM cell viability was detected (Fig. 2b, c).

The obtained data enabled us to conclude that HL1 had earlier and more profound toxic effect as compared to HL2 regardless of leukemic cell lines. Toxic effect of HL2 was detected only at 72 h of incubation of Jurkat and Molt-16 cells (Fig. 2a, b).

The higher HL1 toxic effect could be connected with the presence in its structure of Cl atoms, which possess alkylating potential and are the constituents of chemotherapeutic drug such as cisplatin [29].

The high tumor-specific toxicity of  $\beta$ -diketones derivatives was confirmed by evaluating their effects against human carcinoma cell lines HSC-3 (oral squamous), HSG (submandibular gland), and HL-60 (promyelocytic leukemia) as well [1].

It was shown that  $\beta$ -diketones exist mainly in the enolic form and form metal chelates with Fe, Cu, and Zn ions. Recent studies suggest that metal chelates

induce apoptotic cell death in various tumor cell lines and they are potential antitumor agents against malignant melanomas [30, 31].

To estimate the possibility to enhance the toxicity of studied compounds, their combined action with  $C_{60}$  fullerene on leukemic cells was investigated.

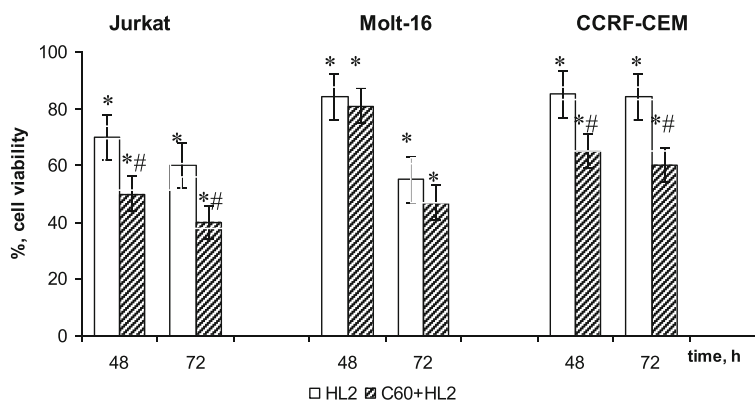
It was shown that  $C_{60}$  fullerene ( $10^{-5}$  M) does not affect the viability of leukemic cells during incubation period (data not shown).

No increase of HL1 toxicity was detected when Jurkat, Molt-16, or CCRF-CEM cells were preincubated with  $C_{60}$  fullerene (data not shown).

At combined action of  $C_{60}$  fullerene and HL2, an enhanced toxic effect in comparison with the effect of HL2 taken separately was observed. In this case, a viability of Jurkat and CCRF-CEM cells was additionally decreased by 20 and 24% at 72 h, respectively (Fig. 3).

At the same time, no enhancement of HL2 toxic effect under the combined action with  $C_{60}$  fullerene against Molt-16 cells was found (Fig. 3).

In vitro experiments combined with modeling simulation have shown that  $C_{60}$  fullerene is significantly accumulated in human leukemic K562 cells



**Fig. 3** Viability of leukemic cells at the combined action of 1 mM HL2 and C<sub>60</sub> fullerene. M ± m, n = 8; \*p < 0.05 compared to control cells; #p < 0.05 compared to HL2

since it could not be efficiently effluxed by P-gp protein [32]. So, C<sub>60</sub>-induced enhancement of HL2 cytotoxicity could be triggered by C<sub>60</sub> fullerene intracellular interactions at the level of membrane phospholipids, cytosolic proteins, DNA, and other biological structures. It is well known that nanoparticles can interact with organic molecules using van der Waals forces, hydrophobicity, π-interactions, and enthalpy driven [33]. Previously with the use of molecular modeling, we have shown that stable C<sub>60</sub>-DNA complex is formed after C<sub>60</sub> fullerene binding with DNA [5].

Computer simulation of HL1 and HL2 interactions with DNA was used to estimate the possible nature of their bonds with biological molecules and the ability of C<sub>60</sub> fullerene to modify such interactions.

It was shown that HL1 forms the stable complexes with DNA when bound with a large groove and during the intercalation into a small groove. So, when the binding shift of DNA double helix was 3.07 Å, and for HL1, it equals to 1.8 Å; at intercalation, the

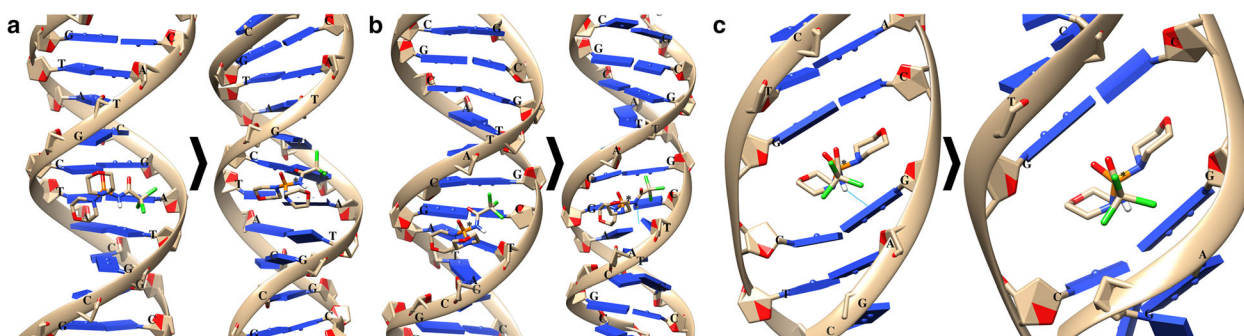
DNA molecule shifted to 3.24 Å, and HL1—2.24 Å (Fig. 4).

According to energy parameters, the formed complexes are not rigid (Table. 1). So, when binding HL1 with a large groove, the contact energy is -34.8 kJ/mol, and at intercalation into DNA, it is -61.0 kJ/mol; the steric clashes between DNA and HL1 in these cases are 1.4 and 7.7 kJ/mol, respectively.

It is shown that HL2 forms the stable complexes with DNA only when bound with a small groove (Fig. 5a). The shift of DNA is 2.09 Å, and for HL2—1.45 Å.

When binding HL1 as well as HL2, observe no changes in their nucleotide environment (only insignificant movements in space take place).

The interaction of the DNA molecule with HL1 or HL2 in combination with C<sub>60</sub> fullerene was studied depending on the sequence of interactions. If initially C<sub>60</sub> fullerene binds with DNA, and then HL1 or HL2, the formed structure is designated as C<sub>60</sub> + HL1 or C<sub>60</sub> + HL2; otherwise—HL1 + C<sub>60</sub> or HL2 + C<sub>60</sub>. We



**Fig. 4** The interaction of DNA molecule with HL1: **a** and **b**—binding with small and large grooves; **c**—intercalation into a small groove. The used DNA structures of the PDB database: **a** and **b**—2M2C; **c**—1XRW

**Table 1** The energy parameters (in kJ/mol) of interaction of the studied structures with DNA double helix

Structure	The energy parameters				
	FreE	Cntc	Hbnd	Bump	Int
The binding with a large groove					
HL1	-1.8	-34.8	-0.4	1.4	4.0
HL2	-3.6	-41.7	-1.8	1.6	3.7
HL1 + C <sub>60</sub>	-20.3	-70.5	-1.1	7.8	3.1
HL2 + C <sub>60</sub>	-20.3	-66.8	0.0	6.5	3.9
The binding with a small groove					
HL1	-4.9	-49.0	-2.7	3.9	4.8
HL2	-4.7	-61.1	0.0	11.2	7.2
HL1 + C <sub>60</sub>	-24.0	-87.8	0.0	11.2	5.3
HL2 + C <sub>60</sub>	-31.1	-84.3	-0.8	6.3	3.8
The intercalation into a small groove					
HL1	-7.0	-61.0	-1.7	7.7	6.8
HL2	2.3	-63.0	0.0	12.6	10.4
HL1 + C <sub>60</sub>	-22.5	-107.0	-1.4	19.6	7.2
C <sub>60</sub> + HL1	-22.8	-97.1	0.0	17.1	4.2
HL2 + C <sub>60</sub>	-21.8	-100.0	0.0	18.6	5.1
C <sub>60</sub> + HL2	-26.5	-89.9	-2.4	12.7	4.2

*FreE* the total energy of binding DNA and related structure, *Cntc* the contact energy of interacting compounds (the related structure with DNA), *Hbnd* the energy of hydrogen interactions, *Bump* the energy of steric clashes between DNA and build-in structure, *Int* the energy of steric clashes between the atoms of build-in structure

considered two options of the binding-studied structures with DNA molecule, namely with small and large grooves, and also the possible intercalation of these structures into the small groove (Figs. 6 and 7). The calculated energy parameters of the interaction of these structures with DNA molecule are given in Table 1.

HL1 + C<sub>60</sub> or C<sub>60</sub> + HL1: at binding HL1 + C<sub>60</sub> structure with small and large grooves of DNA, there is a

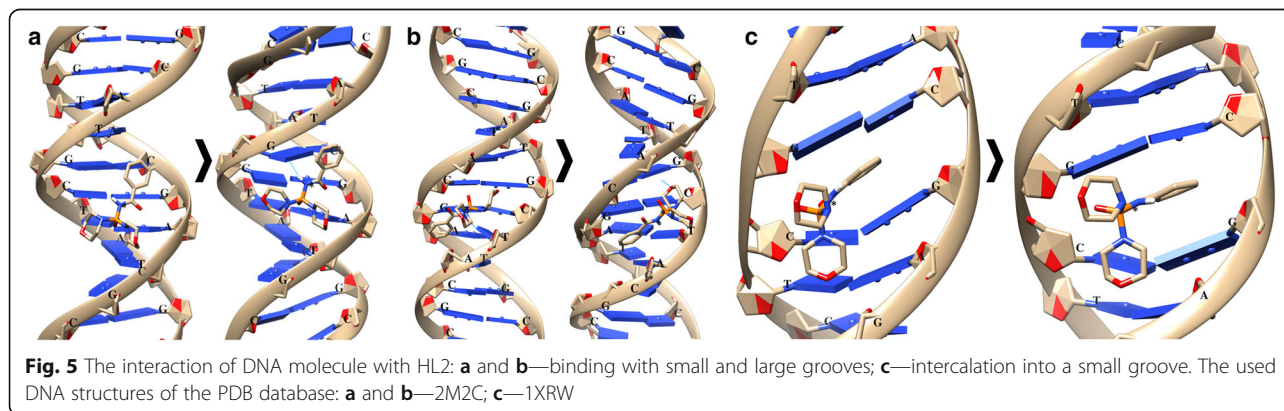
significant shift of the C<sub>60</sub> fullerene compared to HL1 compound: Rmsd value is 5.22 Å in a small groove, and in a large groove—8.08 Å; meanwhile, Rmsd value for HL1 is 2.92 and 2.93 Å in small and large grooves, respectively (Fig. 6a, b).

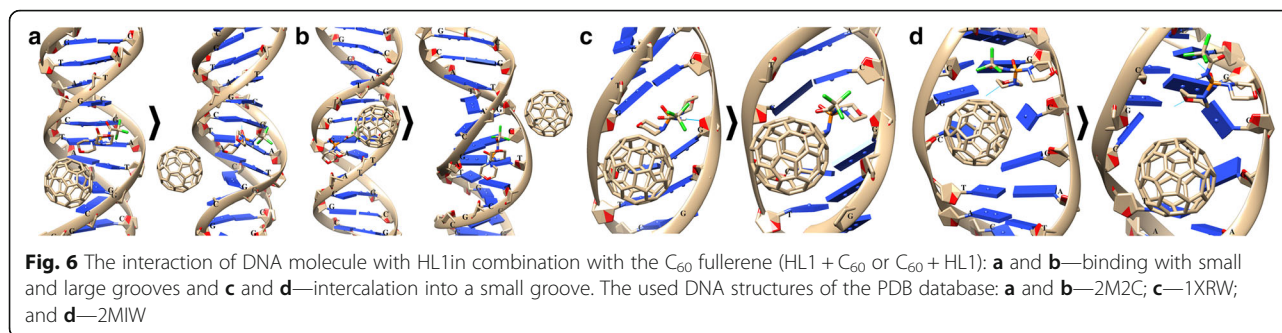
At intercalation of the studied structures into a small groove of the DNA molecule, the significant values of Rmsd for C<sub>60</sub> fullerene in the cases of HL1 + C<sub>60</sub> (5.72 Å) and C<sub>60</sub> + HL1 (6.42 Å) are also observed. Due to such shift of C<sub>60</sub> molecule, a partial displacement of HL1 compound (Rmsd value is 1.99 Å) from the intercalation site (Fig. 6c) takes place. In the case of C<sub>60</sub> + HL1, a similar effect is observed for C<sub>60</sub> molecule (Rmsd value for HMF compound is 2.69 Å) (Fig. 6d).

HL2 + C<sub>60</sub> or C<sub>60</sub> + HL2: in these cases, there is a significant shift of the C<sub>60</sub> fullerene with respect to the HL2 compound at their combined interaction with DNA in small and large grooves (Fig. 7a, b). So, Rmsd value for C<sub>60</sub> fullerene in a small groove is 7.16 Å (for HL2—1.69 Å), and in a large groove—8.56 Å (for HL2—3.85 Å). However, a complete break of stacking interaction between C<sub>60</sub> fullerene and HL2 does not happen.

At intercalation of the studied structures into a small groove of DNA, the C<sub>60</sub> fullerene forms the strong stacking interactions with HL2 and nitrogenous bases of DNA double helix (Fig. 7c, d). So, after molecular docking, its shift is negligible: in the case of HL2 + C<sub>60</sub>, the Rmsd value for C<sub>60</sub> fullerene is 3.99 Å (for HL2—1.36 Å), and in the case of C<sub>60</sub> + HL2—2.7 Å (for HL2—3.29 Å). In addition, at such intercalation, there is no displacement of chemical compound from the original nucleotide environment.

As shown in Table 1, the energy parameters of C<sub>60</sub> + HL1 or C<sub>60</sub> + HL2 intercalation into DNA molecule are lower for C<sub>60</sub> + HL1 structure than for HL1 + C<sub>60</sub> or HL2 + C<sub>60</sub> structures. The Bump value for C<sub>60</sub> + HL1 structure is 17.1 kJ/mol and Int—4.2 kJ/mol, while for





HL1 +  $C_{60}$  structure, these parameters are higher—19.6 and 7.2 kJ/mol, respectively. These data allow to suggest that the studied chemical compounds form more stable structures with  $C_{60}$  fullerene when  $C_{60}$  molecule is initially bound to DNA.

Taking into account the structural features of the studied compounds and  $C_{60}$  fullerene, one can suggest the formation of cation- $\pi$  bonds between HL1 and  $C_{60}$  fullerene due to  $CCl_3$  group (Fig. 6a) and stacking interactions between HL2 and  $C_{60}$  fullerene mediated by the presence of benzene ring (Fig. 7a).

Thus, using the computer modeling, it was found that HL2 and HL1 compounds in combination with  $C_{60}$  fullerene are potentially capable of binding to DNA molecule.

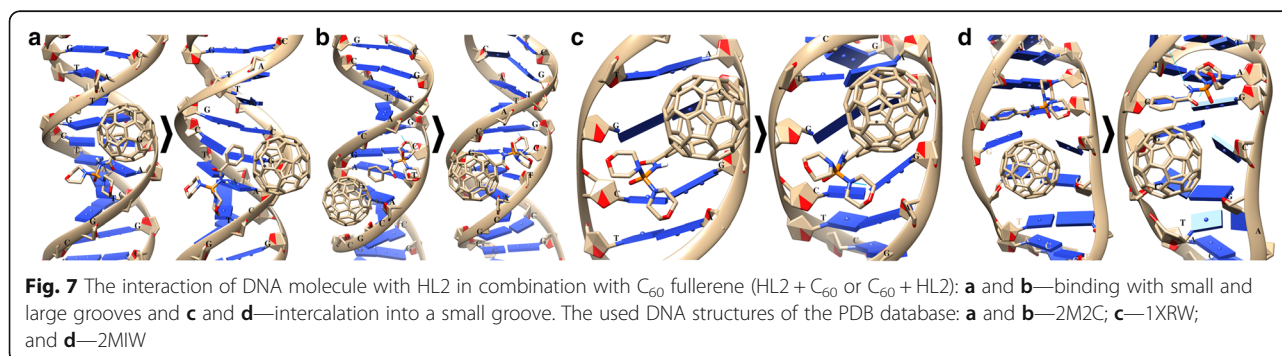
## Conclusions

We have estimated effect of two carbacylamidophosphate derivatives with different substituents on the viability of leukemic cells of three lines. Dimorfolido-*N*-trichloroacetylphosphorylamide (HL1) and dimorfolido-*N*-benzoylphosphorylamide (HL2) at 1 mM concentration in the cellular medium caused the decrease of cell viability, which value was dependent on the cell type and duration of incubation. HL1 and HL2 caused the similar gradual decrease of Jurkat cells viability. HL1 had earlier and more profound

toxic effect as compared to HL2 both on Molt-16 and CCRF-CEM cells. Toxic effect of HL2 was detected only at 72 h of Jurkat and Molt-16 cells incubation.

It was shown that  $C_{60}$  fullerene facilitated the toxic effect of HL2 on Jurkat and CCRF-CEM cells at 48 and 72 h.

By use of computer simulation, the interactions of DNA molecule with HL1 or HL2 separately and in combination with  $C_{60}$  fullerene were studied. The chemical compounds formed the stable complexes with DNA: HL1 when binding with a great groove and at intercalation into a small groove, while HL2 only when bound with a small groove. At the combined action of HL1 or HL2 with  $C_{60}$  fullerene, the  $C_{60}$  + HL2 structure formed the stable complex with DNA when bound with a small groove and at intercalation into it, while the  $C_{60}$  + HL1 structure formed the stable complex with DNA only in one case—when binding with a large groove. In this case, the strong stacking or cation- $\pi$  interactions can be formed between these chemicals and  $C_{60}$  fullerene. Thus, the chemical compounds form the stable complexes with DNA both individually and in combination with  $C_{60}$  fullerene, but the differences in the types of bonds and ways of binding could be the cause of their different cytotoxic effects.



### Abbreviations

DMSO: Dimethyl sulfoxide; DNA: Deoxyribonucleic acid; FBS: Fetal bovine serum; HL1: Dimorfolido-*N*-trichloroacetylphosphorylamide; HL2: Dimorfolido-*N*-benzoylphosphorylamide; IR: Infrared spectroscopy; MTT: 3-(4,5-Dimethyl-2-thiazolyl)-2,5-diphenyl-2-H-tetrazolium bromide; NMR: Nuclear magnetic resonance; RPMI - 1640 medium: Roswell Park Memorial Institute medium

### Authors' Contributions

The work presented here was carried out in collaboration between all the authors. ST, SI, AV, and SM synthesized and characterized dimorfolido-*N*-trichloroacetylphosphorylamide and dimorfolido-*N*-benzoylphosphorylamide. HV and PY performed the computer modeling of interaction of DNA molecule with dimorfolido-*N*-trichloroacetylphosphorylamide or dimorfolido-*N*-benzoylphosphorylamide separately and in combination with C<sub>60</sub> fullerene. GA and FM estimated the viability of leukemic cells. PS, GI, and MO coordinated the experimental work, analyzed the data, performed the statistical analysis, and wrote the manuscript. RU synthesized and characterized C<sub>60</sub> fullerene aqueous solution. All authors discussed the results and commented on the manuscript. All authors read and approved the final manuscript.

### Competing Interests

The authors declare that they have no competing interests.

### Ethics Approval and Consent to Participate

No human participants, tissue, or animals were involved in this study. The work was performed using cell lines from the Collection of Microorganisms and Cell Cultures of the Leibniz Institute DSMZ-Germany.

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