cDNA Microarray Analysis of Adaptive Changes after Renal Ablation in a Sclerosis-Resistant Mouse Strain

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Key Words

Remnant kidney • Renal fibrosis • Transforming growth factor- β_1 • Oxidative stress • Matrix metalloproteinases • ADAMTS • Tissue inhibitors of metalloproteinases

Abstract

Background: 5/6 nephrectomy (Nx) in susceptible animals causes glomerular sclerosis and interstitial fibrosis in the remnant kidney. Oxidative stress, transforming growth factor- β (TGF- β), and the de novo synthesis of collagen seem to contribute to this process. However, these factors might also be required for tissue repair without fibrosis. **Methods:** We examined dynamic changes after nephron loss in a mouse strain capable of complete recovery. C57BL/6 mice underwent single-session Nx and were followed for 40 weeks. Gene expression was monitored over 20 days using 22,000 cDNA microarrays. **Results:** The mice developed transient hypertension and glomerular hypertrophy after Nx but failed to progress to glomerular sclerosis or renal failure. Gene expression profiles revealed three stages of recovery, an early phase of injury response, an intermediate phase of extracellular matrix (ECM) production and a later phase of reconstitution. Surprisingly, oxidative stress responses and

collagen production were strongly upregulated soon after Nx. Furthermore, TGF- β_1 and connective tissue growth factor were rapidly upregulated and remained elevated. **Conclusion:** We suggest that oxidative stress, collagen production, profibrotic growth factors and ECM turnover are part of the comprehensive adaptation to nephron loss and not necessarily associated with progressive loss of renal function.

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Introduction

Progression of chronic renal disease remains a significant clinical problem. A limited renal insult with nephron loss commonly leads to glomerular hyperfiltration, followed by proteinuria, renal fibrosis and end-stage renal disease. Profibrotic growth factors like transforming growth factor- β_1 (TGF- β_1) and connective tissue growth factor (CTGF) play a central role in this process [1]. They stimulate the expression of extracellular matrix (ECM) components, inhibit matrix-degrading enzymes and induce epithelial-mesenchymal transition (EMT), thus promoting renal fibrosis [2]. Release of these growth factors and the subsequent production of collagen have be-

come the central paradigm of progressive renal failure after renal ablation, diabetic nephropathy, and chronic transplant nephropathy.

The 5/6 nephrectomy (Nx) animal model is well established to study progressive renal disease. Most rodent strains are susceptible to renal ablation and undergo a typical course of events. An initial increase in single-nephron glomerular filtration rate leads to glomerular hypertrophy, followed by a decline in renal function and proteinuria within few months. Histology reveals glomerular sclerosis and progressive interstitial fibrosis. In contrast to most rats, several mouse strains recover from Nx without major sequelae [3]. Whereas 129/SV and FVB mice develop at least moderate proteinuria and renal fibrosis after Nx, kidneys of C57BL/6 mice seem hardly affected.

We therefore used C57BL/6 mice to study the beneficial adaptive changes after nephron loss through partial nephrectomy. Besides functional and histological examination we used cDNA microarrays to determine a time course of gene transcription. Even in the absence of subsequent fibrosis or renal failure, we identified an increased transcription of profibrotic growth factors, oxidative stress response and ECM components as part of the healing process.

Subjects and Methods

Animal Preparation

Eight-week-old male C57BL/6 mice were obtained from Charles River (Kisslegg, Germany). The animals were housed at the Animal Research Facility of the University Hospital Freiburg, and had free access to food and water. Body weights were determined at the start of the protocol, the time of surgery, and the time of sacrifice. To synchronize adaptive responses, Nx was performed as a one-step procedure using 8-week-old male mice. After removing the right kidney, the left kidney was decapsulated, and the upper and lower poles were resected under anesthesia with xylazine-ketamine intraperitoneally. Dissected tissues were weighted, and remnant kidney function and weight as well as mean arterial blood pressure were studied at various time points thereafter in 4 animals per time point. Proteinuria after 40 weeks was determined in 3 animals and compared with age-matched controls. Additionally, 3 mice per time point were sacrificed to collect remnant kidneys for RNA extraction as described below. The kidneys obtained from the heminephrectomies were used as the control samples at day 0.

Immunohistochemistry and Electron Microscopy

Remnant kidneys were subject to retrograde perfusion fixation using 3% glutaraldehyde and embedded in paraffin. Four-micrometer sections were cut, deparaffinized and stained with hematoxylin/eosin or periodic acid-Schiff (PAS) to evaluate the

degree of glomerular and tubulointerstitial changes. Immunohistochemistry was performed essentially as described [4]. The washing buffer was PBS containing 0.04% Triton X-100 and 0.35% DMSO. All incubations were performed in a moist chamber at 4°C. Endogenous peroxidase activity was quenched with 0.3% H₂O₂ for 30 min. Anti-MT-1/2 mAb was obtained from DAKO (Hamburg, Germany) and tissues were incubated at 1:100 overnight. M2 anti-FLAG antiserum was obtained from Sigma-Aldrich (Taufkirchen, Germany). For indirect immunoperoxidase staining we used the Vectastain ABC kit (Vector Laboratories, Burlingame, Calif., USA) following the manufacturer's protocol.

For electron microscopy, fixation was performed with 3% glutaraldehyde in 0.08 M cacodylate buffer (pH 7.4) for 2 h. Slices were then rinsed in 0.1 M cacodylate buffer (pH 7.4). The tissue slices were postfixed in 2% OsO₄ in a veronal acetate buffer, and then dehydrated in a graded series of ethanol including en block staining (1 h) in 1% uranyl acetate at the 70% ethanol step. The specimens were then transferred to propylene oxide, and subsequently embedded in Epon-Araldite. Eighty-nanometer sections were cut with diamond knives, mounted on naked grids, and examined with a Philips CM 10 electron microscope.

Glomerular Geometry

Glomerular volumes were determined as described previously [5]. Briefly, kidneys were dissected into 1-μm slices perpendicular to the longitudinal axis. By using area-weighted sampling, 10 small pieces were selected for embedding in Epon-Araldite. Remaining kidney slices were embedded in paraffin, yielding one representative section of each slice for morphometric and stereological investigations. By using systematic subsampling, a Zeiss eyepiece with a 100-point grid (magnification, ×100 and ×400; Integrationsplatte II; Zeiss, Oberkochen, Germany), and the point-counting method ($P_p = A_A = V_V$), the volume densities of cortex, medulla, and glomeruli were determined. The glomerular number per area (N_A) was counted, and the total glomerular volume (V_{glom}) was calculated by multiplying the volume density of glomeruli by the cortex volume ($V_{\text{Vglom}} \times V_C$). The glomerular number per volume (N_V) was determined using the following equation: $N_V = k/\beta \times N_A^{1.5}/V_V^{0.5}$, with $k = 1.03$ and $\beta = 1.382$. The total glomerular number (N_{glom}) was calculated by multiplying the glomerular number per volume (N_V) by the cortex volume (V_C). The mean glomerular volume (v) was then derived from the total glomerular volume and the total number of glomeruli per kidney ($V_{\text{glom}}/N_{\text{glom}}$).

Direct Systolic Blood Pressure Measurements and Urinalysis

Mice were anesthetized with pentobarbital (50 mg/kg, i.p.) and placed on a heated board to maintain body temperature at 37°C. A polyethylene catheter was inserted into the bladder for urine collection. Systolic and diastolic blood pressures were obtained via the left femoral artery by using a strain-gauge blood pressure transducer. Mice were euthanatized after the experiment. Urinalysis was performed colorimetrically using the Bio-Rad Protein Assay (Life Science, Munich, Germany) and the Sigma creatinine kit (Sigma-Aldrich, Munich, Germany).

Microarrays

Remnant kidneys were harvested from 3 animals at days 1, 2, 5, 10, 14 and 20 after Nx, respectively. Gene expression was determined for each remnant kidney in comparison with reference

mRNA, and the logarithmic level of mRNA expression of normal kidneys (day 0) was subtracted for each time point. cDNA microarrays were produced and processed essentially according to the Stanford protocol (<http://cmgm.stanford.edu/pbrown>). All genes were obtained as bacterial stocks from the National Institute of Aging (<http://lgsun.grc.nia.nih.gov/cDNA/15k.html>, http://lgsun.grc.nia.nih.gov/cDNA/NIA_7_4k.html), kindly distributed by the MRC, Cambridge, UK [6, 7]. Plasmids were purified using the Qiagen 96-well Turbo Kit (Qiagen, Hilden, Germany), and inserts were purified by PCR using vector primers flanking the individual inserts (5'-CTG CAA GGC GAT TAA GTT GGG TAA C-3' and 5'-GTG AGC GGA TAA CAA TTT CAC ACA GGA AAC AGC-3'). PCR products were purified by ethanol precipitation, and resuspended in H₂O. Aliquots were dried and resuspended in 3× SSC to a final concentration of approximately 40 ng/μl. Printing was performed on aminosilane-coated slides (CMT-GAP II Slides, Corning, N.Y., USA), using an arrayer that was assembled according to specifications by the Stanford group using software kindly provided by Joe de Risi (<http://cmgm.stanford.edu/pbrown>).

RNA Preparation

Frozen tissues were homogenized in 4 M guanidinium isothiocyanate with 0.72% β-mercaptoethanol using a polytron (PT-MR2100, Kinematica, Lucerne, Switzerland). Total RNA was subsequently purified on a cesium chloride gradient. After ethanol precipitation RNA was resuspended at a concentration of 0.4–4 μg/μl, and stored at –80°C.

Hybridization

All hybridizations were performed in the presence of an equal amount of reference RNA (Stratagene, La Jolla, Calif., USA) as published [8]. Twenty-four micrograms of either tissue or reference RNA were transcribed into cDNA in the presence of Cy3- or Cy5-labeled dUTP, using Superscript II reverse transcriptase (Invitrogen, Carlsbad, Calif., USA). All other steps, including hybridization, were performed following the protocol published by P. Brown et al. (<http://cmgm.stanford.edu/pbrown>). A PCR purification kit (Qiagen) was used for cDNA purification.

Semiquantitative RT-PCR

Total RNA (1 μg) from two remnant kidneys was purified for each time point with the RNase-free DNase set (Qiagen) and reversely transcribed into cDNAs using oligo (dT) 12–18 primers and Superscript II reverse transcriptase (Invitrogen). Each PCR was carried out with 1,4-β-galactosidase as internal control in each reaction vial. The following primers were used (5'–3'): β-galactosidase (BG071892): caggcagctcgattgaaaa, caaggtgcacactcctgta; TGF-β₁: gtcagacattcgggaagcag, ctgccgtacaactccagtga; CTGF: caaagcagctgcaaatacca, ggccaaatgtgtcttccagt. The PCR was initiated by denaturation (94°C, 5 min), followed by 28 cycles of denaturation (30 s), annealing (1 min, 55°C), and elongation (2 min, 72°C). PCR products were run on 3% agarose gels, and evaluated in relation to the corresponding β-galactosidase band using Scion Image software (Scion, Frederick, Md., USA). Array results were confirmed by RT-PCR for the following genes (data not shown): collagen 3A1 (BG086357), SPARC (BG064802), heme oxygenase-1 (HO-1; BG077732), FN14 (BG080700), and isopentyl-diphosphate delta-isomerase (BG079889).

Data Analysis

Signal intensities were measured by an Axon 4000A scanner using GenePix 3.0 software (Axon Instruments, Union City, Calif., USA). Image and data files, array layout, and all relevant information according to the MIAME guidelines (Minimum Information about a Microarray Experiment) were transferred to the GeneTrafficDuo database (Microarray Data Management and Analysis Software, Iobion Informatics, La Jolla, Calif., USA). To exclude artifacts near background range, all spots were eliminated when sample intensity or reference intensity was less than 50 or less than the local background. Local background was subtracted from spot intensities. Normalization was performed by the locally weighted scatter plot smoother subgrid normalization method. Log ratios of each gene are transformed to a mean of zero. An analysis of variances (ANOVA) was applied to detect differentially expressed genes. The p values obtained were controlled for multiple testing (FDR = false discovery rate). An adjusted p value smaller than 0.05 was considered significant. Agglomerative hierarchical clustering was performed using the R statistical software package (www.r-project.org). Correlation was used to measure the distances of genes or arrays. Gene expression time courses are grouped by k-means clustering to find main patterns of regulation. To test for enrichment of Gene Ontology (GO) categories [9], the number of appearances of all GO functions of a group of interesting genes is compared with the number of appearances in a reference group. Fisher's exact test was applied and calculated p values are corrected for multiple testing.

Results

Nx in C57BL/6 Mice Is Not Associated with Fibrosis, Glomerular Sclerosis or Proteinuria

Figure 1 demonstrates the functional changes after Nx in C57BL/6 mice. The remnant kidneys rapidly enlarge, reaching a maximum weight after 9 weeks (fig. 1a). An increase in both systolic and diastolic blood pressure occurred after 1 week and was followed by a decline to the initial range within 2–4 months (fig. 1b). Mean glomerular volumes increased significantly after 1 week with a decline to baseline levels 9 weeks after Nx (fig. 1c). Similarly, urine flow was almost doubled 1 week after renal ablation, but declined to control levels after 16 weeks (fig. 1d). The gain in body weight was not different between operated and control animals. None of the animals developed proteinuria or renal failure. As demonstrated in figure 1e, urinary protein excretion in 3 mice examined 40 weeks after Nx was not different from age-matched controls. Light and electron microscopy performed at various time points up to 40 weeks after Nx revealed occasional mild mesangial hypercellularity but no interstitial renal fibrosis or glomerular sclerosis (fig. 2a–c).

Fig. 1. Functional changes in C57BL/6 mice after Nx. **a** Remnant kidney weight increased rapidly after Nx, peaking after 9 weeks. **b** There was a transient rise in blood pressure. However, both systolic and diastolic blood pressures (BP) approached baseline levels few weeks after Nx. **c, d** Mean glomerular volume and urine flow

increased significantly after 1 week, and normalized within 9–16 weeks. **e** There was no difference in urinary protein excretion between mice 40 weeks after Nx and age-matched controls. Data are presented as mean $\pm$ standard deviation.

Transcriptional Analysis Defines Three Phases of Tissue Response

After selecting 176 significant genes by ANOVA all samples cluster according to their day of retrieval (fig. 3a). There are two major branches, one containing the samples from day 1 and day 2 and the second containing all the others. The latter has three branches. One is made up of controls (day 0) only, one exclusively contains day 5 samples and the third comprises days 10, 14 and 20.

A Cluster of ECM Encoding Genes Shows Peak Expression on Day 5

Of the 176 genes identified by ANOVA to show a significant regulation over time after Nx, 52 had peak expression levels on day 5 (fig. 3b). We suspected day 5 to be a turning point between early acute-phase response and

late successful reconstitution. Therefore, gene ontology analysis was performed comparing these 52 genes with the entire array. Among functional groups significantly overrepresented were ECM constituents, collagens and cell adhesion molecules (table 1).

Transcription of Metalloproteinases and Their Inhibitors

Matrix accumulation is controlled by extracellular proteinases, in particular matrix metalloproteinases (MMP) and ADAMTS (a disintegrin and metalloproteinase with thrombospondin motifs), and by their inhibitors tissue inhibitors of metalloproteinases (TIMP). As demonstrated in figure 4a, collagen expression increased up to 28-fold after Nx. Maximal levels were reached on day 5. Figure 4b demonstrates a significant upregulation of

Fig. 2. Changes in renal histology after Nx in C57BL/6. **a** Glutaraldehyde-fixed paraffin sections from remnant kidneys. The glomeruli showed a mild degree of mesangial hypercellularity after 9 weeks; the tubulointerstitial compartment appears unchanged. HE. $\times 120$. **b** Slides of remnant kidneys 9 months after Nx and age-matched controls. There was no obvious increase in ECM deposition in the remnant kidney. PAS stain. Original magnification $\times 120$. **c** Electron micrograph of a remnant kidney 9 months after Nx. Both basement membrane and podocyte foot processes were completely normal without any changes suggestive of glomerulosclerosis. Original magnification $\times 27,500$.

MMP-2, MMP-7 and MMP-23, whereas MMP-11 and MMP-9 transcription was not significantly altered after Nx. The MMP inhibitors TIMP-2 and TIMP-4 are also upregulated after Nx peaking on days 5 and 2, respectively (fig. 4d).

ADAMTS-1 and ADAMTS-4 were strongly upregulated (fig. 4c), accompanied by a profound decrease in the transcription of their main inhibitor TIMP-3 (fig. 4d). A mild increase was evident for ADAMTS-2, ADAMTS-10 and ADAMTS-12. ADAMTS-19 transcription, on the other hand, was downregulated after Nx (fig. 4c).

Early and Sustained Upregulation of TGF- β_1 and CTGF

TGF- β_1 has been regarded as the key mediator of fibrogenesis with pleiotropic effects leading to ECM accumulation. Semiquantitative RT-PCR revealed that TGF- β_1 is rapidly upregulated after Nx. TGF- β_1 expression reached a maximum on day 5 and remained elevated (fig. 5a, b). The expression of CTGF, another potentially

profibrotic factor, followed a similar time course. In addition, we noted a prolonged upregulation of proteins that regulate TGF- β_1 action, including the cell surface TGF- β -binding protein betaglycan, decorin and biglycan (fig. 5c).

Oxidative Stress Responses

Among the phase 2 enzymes, the expression of microsomal glutathione S-transferase 1 (mGST1) and glutathione S-transferase omega 1 (GSTo1) were significantly upregulated reaching a maximum at day 1 (fig. 6a). Glutathione synthetase (GSS), on the other hand, was downregulated. Transcription of the other glutathione transferases and synthetases examined was not significantly altered after Nx. HO-1, the rate-limiting enzyme in heme catabolism, was dramatically upregulated, reaching an 18-fold maximum 24 h after renal ablation (fig. 6b). As expected, the expression of the constitutively expressed heme oxygenase-2 (HO-2) remained unchanged. There was also an early upregulation of aldo-keto reductase

Fig. 3. Cluster analysis of transcriptional changes over 20 days after Nx. **a** Agglomerative hierarchical cluster analysis of individual arrays, based on an ANOVA on a significance level of <0.05 . 176 genes fulfilled the criteria and are shown in columns, samples are sorted in rows. Light shades represent upregulation, shades of dark downregulation. Arrays cluster according to the day of retrieval (d). Four larger groups can be observed (day 0, days 1–2, day 5, and days 10–20). **b** Pattern of genes maximally regulated on day 5 after Nx. Time course of 52 significantly regulated genes with a maximum expression on day 5 after Nx. The black line indicates the average expression pattern of these genes, vertical bars indicate standard deviations.

family 1, member B8 (Akr1b8), whereas other members of this family were downregulated.

Cu-Zn superoxide dismutase (SOD) 1 and 2 were not significantly regulated after renal ablation (fig. 6c). In contrast, glutathione reductase (GSR) and thioredoxin reductase (Txnrd1) were increased. Glutathione peroxidase Gpx1 decreased, reaching a minimum at day 2 after Nx. Metallothionein-2 (MT-2) was among the antioxidants regulated most significantly reaching a 12-fold increase shortly after renal ablation (fig. 6d). Interestingly, leukemia inhibitory factor (LIF), a factor involved in mesenchymal-to-epithelial conversion of renal tubular cells and known to induce MT-2, was also upregulated after Nx, closely following the time course of MT-2 (fig. 6d). The increase in MT-2 transcription was associated with increased translation as shown by immunohistochemistry (fig. 6e).

Discussion

Transcriptome analysis using microarray technology represents a powerful tool to study mechanisms of disease as well as physiologic adaptive changes. This technique has been used in numerous mouse models of progressive renal disease and helped to identify profibrotic factors and prognostic markers. However, some of these factors might also be involved in the healing process after kidney injury. To clarify this, we addressed the dynamic response in a mouse strain capable of recovering from Nx without developing renal fibrosis or proteinuria.

In our C57BL/6 mice, there was a gain in remnant kidney weight after Nx peaking around week 9. Similar findings have been described in other mouse and rat models and are attributed to renal hyperplasia and hypertrophy of the remaining nephrons. Systemic blood pressure increased temporarily but reached baseline levels within 9 weeks. A rise in blood pressure after Nx is typical, and attributed to the activation of the renal renin-angiotensin-aldosterone system. However, the subsequent nor-

Table 1. Gene ontology functional categories most significantly overrepresented on day 5 after Nx

Gene ontology category	p FDR
ECM (sensu Metazoa)	0.000000
ECM structural constituent (tensile strength)	0.000000
Collagen	0.000000
ECM structural constituent	0.000000
Extracellular space	0.000000
Phosphate transport	0.000000
Structural molecule activity	0.000002
Cell adhesion	0.000028
Calcium ion binding	0.001000
Dipeptidyl-peptidase I activity	0.001173
Positive regulation of type IIa hypersensitivity	0.001596
Lysosome	0.001716
Mitotic spindle checkpoint	0.001887
Positive regulation of phagocytosis	0.002979
Actin binding	0.002979
Basement membrane	0.004667
Complement activation	0.007108
Membrane	0.009607
Cytoplasm	0.016338
Calmodulin binding	0.016410

Of the 176 genes identified by ANOVA to show a significant regulation over time after Nx, 52 had peak expression levels on day 5. These 52 genes were tested for enrichment of GO categories in relation to the entire array. The 20 functional categories most significantly overrepresented in this group and their adjusted p values are listed.

FDR = False discovery rate.

malization in this mouse strain is rather unusual. Furthermore, we did not observe signs of renal fibrosis or glomerulosclerosis by light microscopy or electron microscopy within 40 weeks after Nx, and there was no development of proteinuria. The absence of fibrotic changes confirms other studies suggesting that C57BL/6 mice do not develop long-term sequelae after subtotal renal ablation [3].

Using ANOVA and cluster analysis of differentially expressed genes of remnant kidneys collected during 20 days after Nx, we found that samples clustered according to their day of retrieval. This finding indicates a universal response without major interindividual differences. The dendrogram supports the existence of three distinct phases in the adaptive response to Nx. An early acute-phase reaction (days 1 and 2) is followed by a distinct intermediate state (around day 5) and a later phase of reconstitution (days 10–20). Transcription peaking on day 5 included mainly genes encoding ECM components such

as collagens or fibronectin. Interestingly, they are down-regulated again towards day 20. This may be indicative of transient matrix production as required for tissue repair without excessive scarring or progressive fibrosis.

Abnormal ECM accumulation is a common finding in renal fibrosis, and altered matrix degradation is regarded as a key factor in this process. However, ECM deposition and turnover may also be beneficial and essential after tissue injury. We observed a delayed upregulation of MMP-2 transcription, a secreted metalloproteinase with pleiotropic effects. MMP-2 is an antifibrotic enzyme facilitating matrix degradation but has also been associated with renal scarring. It can degrade the tubular basement membrane thereby inducing EMT with resultant tubular atrophy and fibrosis [10]. Interestingly, we observed an upregulation of TIMP-2 transcription with an almost identical time course, suggesting that some of the functions of MMP-2 might be blocked locally on a posttranslational level. MMP-7 was also significantly upregulated during the entire study period of 20 days after Nx. While the gene is hardly expressed in normal kidneys, an up-regulation has been observed in various models of irreversible renal fibrosis [11]. MMP-7 can cleave E-cadherin thereby facilitating epithelial reorganization and possibly EMT. In fact, increased levels of MMP-7 mRNA have been identified as a marker of poor prognosis in human hydronephrotic kidneys [12]. Nevertheless, our data suggest that this MMP might also have beneficial effects in tissue repair.

Transcription of the metalloproteinases ADAMTS-1 and ADAMTS-4 was strongly upregulated peaking on day 1 after Nx. These enzymes are mainly responsible for degrading aggrecan, a process involved in cartilage destruction as seen in arthritides. However, aggrecan can also protect collagen fibrils from degradation by collagenases [13], so the function of these ADAMTS in our setting might be to facilitate collagen turnover. Interestingly, the gene of their main inhibitor TIMP-3 was inversely regulated, possibly enhancing their effect on ECM degradation.

The growth factors TGF- β_1 and CTGF have been identified as key players in the development of renal fibrosis in various human diseases and animal models. Surprisingly, we also found a profound upregulation of their transcription in our model of successful recovery from nephron loss. Although ectopic expression of TGF- β_1 causes renal fibrosis [14], TGF- β_1 is a pleiotropic growth factor and the cellular effects may depend on its spatial and temporal activation. Interestingly, TGF- β_1 expression after Nx was accompanied by a prolonged upregula-

Fig. 4. Transcriptional changes in metalloproteinases and their inhibitors after Nx. **a** Collagen expression increased profoundly after Nx reaching a maximum on day 5. Collagen type III alpha 1 (Col3a1) showed the highest expression level (30-fold). **b** Upregulation of MMP. Expression of MMP-2 and MMP-23 is upregulated after Nx peaking on day 5 and remains increased throughout the study period. MMP-7 is also increased peaking on days 2 and 14 after Nx. **c** Changes in ADAMTS. There is a significant

early upregulation of ADAMTS-1 and ADAMTS-4 (day 1), followed by an increase in ADAMTS-10 (days 5–20). Expression of ADAMTS-19, on the other hand, is slightly downregulated. **d** Changes in TIMP. Whereas TIMP-2 and TIMP-4 were upregulated peaking on days 5 and 2, respectively, TIMP-3 was downregulated early after Nx and remained low throughout the study period.

tion of proteins that regulate its action, such as betaglycan, decorin, and biglycan. Betaglycan has been reported to inhibit TGF- β_1 signaling in renal cells by preventing the dimerization of TGF- β receptors, and both decorin and biglycan have been demonstrated to inactivate TGF- β_1 [15]. In addition, decorin can lower glomerular levels of TGF- β_1 , and ameliorate ECM accumulation [16]. Thus, expression of TGF- β_1 alone does not necessarily predict or cause renal fibrosis, but may require the permissive effects of regulatory partners to inflict renal damage.

CTGF mRNA expression is increased in tubular epithelial cells in a variety of chronic renal diseases and has been regarded a promising target of antifibrotic therapy [1]. In subtotaly nephrectomized TGF- β_1 transgenic

mice the intravenous administration of CTGF antisense oligodeoxynucleotide from day 29 to 56 significantly reduced levels of CTGF mRNA in the tubular epithelium and limited renal interstitial fibrogenesis [2]. Our data, however, suggest that an early upregulation of this gene is not necessarily associated with renal fibrosis but might also be required for tissue repair.

Oxidative stress during the early adaptive phase after Nx has been implicated in the pathogenesis of long-term glomerular sclerosis, and therapy with antioxidants can ameliorate the course of renal failure after renal ablation [17]. Reactive oxygen species produced by an increased metabolism and oxidative phosphorylation after renal injury triggers TGF- β , thereby providing a direct link be-

Fig. 5. Upregulation of profibrotic growth factors after Nx. **a** RT-PCR products for TGF- β_1 and CTGF genes. PCR was performed on RNA from 2 animals for each gene and time point. β -Galactosidase (β -gal), a gene not differentially regulated after Nx, was used as an internal control. One example for each PCR reaction is shown; the lanes correspond to day 0, 1, 2, 5, 10, 14, 20 (from left to right in duplicate). **b** Normalization of the data shown in **a** demonstrates increased levels of both growth factors after Nx. Expression of both genes showed an early peak (CTGF on day 1, TGF- β_1 on days 2–5), and slightly decreased again towards day 20. The

graphic depicts the ratios of gene of interest and β -galactosidase over the time course (medians of the duplicates from one experiment are shown). **c** Upregulation of small leucine-rich proteoglycans and betaglycan after Nx. Relative changes of mRNA expression were determined by cDNA microarray. The TGF- β_1 -binding proteins decorin, biglycan and betaglycan were strongly upregulated after renal ablation. Changes were most pronounced for decorin, revealing a 4-fold increase. The levels of all three genes remain elevated throughout the observation period.

tween reactive oxygen species generation and renal fibrosis. Our analysis revealed that Nx induces a complex expression pattern of antioxidants. HO-1, MT-2, mGST1, GSTo1 and Txnrd1 were rapidly upregulated (day 1), followed by Akrlb8, GSR (day 2) and Gpx3 (day 5). Expression of GSS, Akrla4 and Gpx1, however, decreased early after Nx. HO-1 is triggered by several stimuli, including heme, hydrogen, cytokines, and growth factors. However, the precise molecular mechanism of its induction after Nx remains to be elucidated. It is interesting to note that dopamine triggers HO-1 in vitro, and favorably affects

the outcome of cadaveric kidney transplantation [18]. Thus, it is possible that the rapid increase of HO-1 after Nx is mediated by adrenergic stimuli and/or changes in hemodynamics. Glutathione peroxidases are selenium-containing enzymes that catalyze the reduction of various hydroperoxides. A Gpx-like protein has been shown to represent a H_2O_2 sensor in *Saccharomyces cerevisiae* necessary for hydrogen peroxide homeostasis [19]. Hence, the generation of reactive oxygen species may act as a second messenger that induces a comprehensive antioxidant program to prevent further damage. MT-2 belongs to a

Fig. 6. Upregulation of antioxidant systems after Nx. **a** Upregulation of glutathione transferases. Expression of the mGST1 and the GSTo1 increased while other glutathione transferases and synthetases were basically unchanged or downregulated [GSS; GSTa4 (glutathione S-transferase alpha 4)]. **b** Upregulation of phase 2 enzymes after Nx. HO-1, but not HO-2, was rapidly upregulated after Nx and remained elevated until day 5. Akr1b8 followed a similar course, whereas Akr1a4 was downregulated. The other phase 2 enzymes examined [UGT (UDP-glucuronosyltransferase); Ephx (epoxide hydrolase)] were unchanged or slightly downregulated after Nx. **c** Upregulation of other antioxidant systems

after Nx. SOD1 or SOD2 were not significantly regulated. In contrast, the GSR and Txnrd1 as well as the glutathione peroxidase Gpx3 were upregulated. It is interesting to note that Gpx3 expression peaks at day 5 after an early decrease. Gpx1 decreased most significantly (day 2). **d** Upregulation of MT-2 and LIF after Nx. Expression of both MT-2 and LIF is rapidly upregulated after Nx and appears to follow a similar time course. **e** Anti-MT-2 staining in remnant kidneys on days 0, 7 and 63. MT-2 expression is seen as dark staining in the tubular epithelium. M2 anti-FLAG anti-serum served as negative control (right, original magnification $\times 100$).

superfamily of intracellular metal-binding proteins with unique redox capabilities. MT-2 is induced by zinc, hypoxia, and various cytokines, and accumulates in rapidly proliferating cells [20]. Interestingly, LIF, a factor involved in mesenchymal-to-epithelial conversion of tubular epithelial cells, closely follows the time course of MT-

2. Taken together, the sequential activation of different antioxidant systems suggests that Nx triggers a well-orchestrated cascade of signaling events that limit the damage caused by reactive oxygen species.

Our results show that oxidative stress responses, increased expression of TGF- β_1 and collagen production

might be part of necessary adaptive mechanisms to subtotal renal ablation. Hence, even though the exact role of single genes and the influence of the genetic background and confounding factors such as hypertension remain elusive, they suggest that these factors are not unconditionally profibrotic but may be counteracted or might even be essential components of successful tissue repair.

These observations question several central paradigms of renal fibrosis: neither oxidative stress responses nor collagen production are necessarily associated with renal fibrosis. Most importantly, our findings suggest

that an early increase in the expression of TGF- β_1 , CTGF, MMPs or ECM components cannot be used to predict renal fibrosis or progressive renal failure in this mouse model.

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