





Article

Combining Isotope Dilution and Standard Addition—Elemental Analysis in Complex Samples

Christine Brauckmann ¹ , Axel Pramann ^{1,*} , Olaf Rienitz ¹ , Alexander Schulze ¹,
Pranee Phukphatthanachai ^{2,3} and Jochen Vogl ² 

¹ Physikalisch-Technische Bundesanstalt (PTB), Bundesallee 100, 38116 Braunschweig, Germany; christine.brauckmann@ptb.de (C.B.); olaf.rienitz@ptb.de (O.R.); alexander.schulze@ptb.de (A.S.)

² Bundesanstalt für Materialforschung und-Prüfung (BAM), Richard-Willstätter-Str. 11, 12489 Berlin, Germany; pranee@nimt.or.th (P.P.); jochen.vogl@bam.de (J.V.)

³ National Institute of Metrology (Thailand) (NIMT), 3/4-5 Moo 3, Klong 5, Klong Luang, Pathumthani 12120, Thailand

* Correspondence: axel.pramann@ptb.de; Tel.: +49-531-592-3219

Abstract: A new method combining isotope dilution mass spectrometry (IDMS) and standard addition has been developed to determine the mass fractions w of different elements in complex matrices: (a) silicon in aqueous tetramethylammonium hydroxide (TMAH), (b) sulfur in biodiesel fuel, and (c) iron bound to transferrin in human serum. All measurements were carried out using inductively coupled plasma mass spectrometry (ICP–MS). The method requires the gravimetric preparation of several blends (b_i)—each consisting of roughly the same masses ($m_{x,i}$) of the sample solution (x) and $m_{y,i}$ of a spike solution (y) plus different masses ($m_{z,i}$) of a reference solution (z). Only these masses and the isotope ratios ($R_{b,i}$) in the blends and reference and spike solutions have to be measured. The derivation of the underlying equations based on linear regression is presented and compared to a related concept reported by Pagliano and Meija. The uncertainties achievable, e.g., in the case of the Si blank in extremely pure TMAH of $u_{\text{rel}}(w(\text{Si})) = 90\%$ (linear regression method, this work) and $u_{\text{rel}}(w(\text{Si})) = 150\%$ (the method reported by Pagliano and Meija) seem to suggest better applicability of the new method in practical use due to the higher robustness of regression analysis.

Keywords: isotope dilution mass spectrometry; standard addition; ICP–MS; blank characterization; silicon; sulfur; transferrin; tetramethylammonium hydroxide; biodiesel fuel; human serum



Citation: Brauckmann, C.; Pramann, A.; Rienitz, O.; Schulze, A.; Phukphatthanachai, P.; Vogl, J. Combining Isotope Dilution and Standard Addition—Elemental Analysis in Complex Samples. *Molecules* **2021**, *26*, 2649. <https://doi.org/10.3390/molecules26092649>

Academic Editor: Zikri Arslan

Received: 20 April 2021

Accepted: 28 April 2021

Published: 30 April 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Combined developments in inductively coupled plasma mass spectrometry (ICP–MS) techniques and instrumentation, with the continuous improvement of the calibration and evaluation of ICP–MS measurements, are currently considered the most accurate and precise tools in metrology in chemistry. This combination enables a remarkable decrease in measurement uncertainties in elemental analysis [1,2]. Related areas of analytical chemistry are geochemistry, environmental sciences, clinical chemistry, and forensics as well as fundamental metrology, to mention just a few [2–7]. Depending on sample composition (e.g., number of isotopes of the element of interest), concentration range, availability, experience of the analyst, availability of certified reference materials (CRMs), intended uncertainty, and other reasons, the determination of element content (in the following: mass fraction w) of a sample (x) can be conducted by a variety of calibration strategies [2]. Beneath simple external one-point calibration options with external standards [8], additional sophisticated methods have been developed. Among them, isotope dilution mass spectrometry (IDMS) concepts are usually the methods of choice, provided that the analyte element consists of at least two stable isotopes. The IDMS approach is considered a primary method in metrology in chemistry: it is fully understood and operated on the highest technological level. Moreover, it yields the smallest measurement uncertainties possible (most accurate

and precise) according to the *Guide to the Expression of Uncertainty in Measurement* (GUM) [9]. Additionally, loss of the analyte after addition and equilibration of the spike does not affect the results. This method, based on the measurement of isotope ratios (intensity ratios), has been successfully applied in analytical chemistry for several decades and is still subject to further improvement [2,10–13]. Several variations of the IDMS method are known, based on the classical “single” IDMS approach. This needs a fully characterized spike material (same element as the analyte, with preferably inverted isotopic composition and known molar mass and purity) for the preparation of a blend consisting of the sample and the spike. However, often, the characterization of the spike is difficult or impossible, leading to “double” and higher-order (“triple”) IDMS approaches [2,14,15]. In “double” IDMS, a second blend is prepared from the spike and a well-characterized reference material, rendering the knowledge of purity and the molar mass of the spike obsolete. In “triple” IDMS, three blends are applied, with the additional advantage that the isotope ratio in the initial spike does not need to be measured. These higher-order approaches yield lower uncertainties at the cost of considerably more effort in sample preparation.

In the case of a monoisotopic system, the standard addition method is usually applied as an internal calibration method, being more sensitive and accurate than any external method [16–18]. This approach is chosen when the analyte is present in a complex matrix and/or a low concentration range. The regression analysis of the linear curve of the measured signal in several aliquots versus the ratio of the masses of the standard and sample aliquot yields in the x -axis intercept the desired mass fraction of the analyte element. A further-developed method is the standard addition method with an internal standard, useful in the case of long-term drifts and matrix effects [8,16]. This method requires the absence of the element of the internal standard in the sample, blank, and calibration standard. In most cases, the sample exists in a more or less different—and sometimes complex—matrix compared to the spike material.

Recently, Pagliano and Meija pioneered this problem in a sophisticated theoretical way by merging isotope dilution and standard addition methods [1]. They aimed at reducing or even circumventing matrix effects due to the matched matrix in all the measured solutions.

In this study, a similar and related method combining isotope dilution and standard addition methods has been developed independently. The intention of this work is to (1) reduce the impact of matrix effects in isotope dilution by combining it with a robust standard addition procedure, (2) simplify and linearize the working equations for the measurements of analyte mass fractions via linear regression to enable an uncertainty analysis according to GUM, (3) enable the determination of even trace amounts in, e.g., blank solutions, and (4) provide isotope ratio measurements with no need to correct for mass bias, which is a real benefit to the practical analyst. The method presented in the current work is demonstrated and validated experimentally using three completely different matrices in elemental analysis using ICP–MS: the determination of silicon with natural isotopic composition in an aqueous TMAH blank solution, the measurement of sulfur in biodiesel fuel (BDF), and the quantification of transferrin (TRF) in human serum.

A special need for the exact determination of the content of silicon with natural abundance—in aqueous TMAH used as a blank solution—is given in the *Avogadro Project* [19]. This was a multidisciplinary approach, initially set up for the realization of the Avogadro constant N_A with the lowest associated measurement uncertainty and, after the SI revision in 2019, aimed at the dissemination of the SI units of kilogram and mole [19–22]. In that context, chemically highly pure silicon, extensively enriched in ^{28}Si , with $x(^{28}\text{Si}) > 0.9999$ mol/mol, was used. For the determination of the respective molar mass M of the silicon, $u_{\text{rel}}(M)$ must be smaller than 1×10^{-8} . This can only be achieved by reducing both the contamination and the quantification of the remaining natural silicon, which is described as an application of the new method presented.

The second example of the new combined method is the measurement of the mass fraction (w_x) of sulfur in a biodiesel fuel matrix. These measurements were performed within an interlaboratory key comparison conducted by the IAWG of the CCQM (Inor-

organic Analysis Working Group of the Consultative Committee for Amount of Substance: Metrology in Chemistry and Biology): CCQM-K123 [23]. The mass fractions of several trace elements (impurities)—among them, sulfur—were determined. Originally, w_x was determined using an established IDMS technique. In addition, one of the biodiesel fuel samples was measured using the new combined IDMS-Standard Addition approach as a complementary method (this work). This enables the comparability of the new approach with an established and validated method. The biodiesel fuel matrix is an extremely complex matrix because of its volatility and impurities. Sulfur occurs in different compounds in biodiesel fuel, and, for obtaining accurate results, the conversion to sulfate must be ensured in the digestion step. Additionally, sulfur contamination during the whole analytical procedure is an issue and must be controlled.

In the third part, the quantification of TRF in human serum is presented as an example for speciation analysis with a clinical background. The origin of this work was a doctoral thesis followed by the EMRP HLT05 project, which was a European metrology research program [15]. TRF is a biomarker for congenital disorders of glycosylation as well as certain cancers and alcohol abuse, and, therefore, determining its concentration in blood aids in effective clinical diagnosis [24]. Due to this, TRF is listed as an important clinical analyte in the Guidelines of the German Medical Association of 2019 (Rili-BÄK) and is routinely measured in clinical laboratories [25] even though the target values are method-dependent since no reference method is available [25]. In clinical laboratories, the common way of quantifying TRF in serum is immune-based methods like immunoturbidimetry [26].

In order to quantify the protein TRF via its iron content, high-performance liquid chromatography (HPLC) was coupled to an ICP-MS. This approach enables us to separate TRF from the complex blood matrix and identify TRF by its characteristic retention time. To this day, the quantification of TRF is performed by applying double or triple IDMS and considering the iron background and complex matrix of the samples [15,27]. The new approach presented in this publication, combining IDMS with standard addition, is used for the first time in a complex biological sample as proof of principle. Since the sample is a certified serum material, the results of the new approach are compared with the certified target value and its reported range.

A central part of this work is also the comparison of the new method presented here with the one reported in [1]. Both methods have the same or a similar intention: the improvement of measurement design and the reduction of uncertainty for the determination of mass fractions w_x of elements in complex matrices. For better understanding and adaption to our notation, we have additionally derived the method reported in [1] from scratch for the three blends case: we ended up with exactly the same equation (which is Equation (9) in [1]). For the comparison, we evaluated all measurements using our new method (this work) and the one reported in [1].

2. Theoretical Methods

The main advantage of the combination of IDMS and standard addition is its unique simplicity of using gravimetrically prepared blends, ensuring traceability to the International System of Units (SI). It is based on the measurement of virtually only one isotope ratio and three masses per blend. The new approach yields, finally, Equation (1) to describe linear regression. The regression parameters, slope a_1 and y -intercept a_0 , are then used to calculate the aimed-at mass fraction w_x of the analyte element in the original sample (x). The detailed derivation of Equations (1) and (2) is given in Appendix A.

$$\underbrace{\frac{m_{y,i}}{m_{x,i}} \times \frac{R_y - R_{b,i}}{R_{b,i} - R_x}}_{=y_i} = \underbrace{\frac{1}{w_y} \frac{M_y \sum R_y}{M_x \sum R_x}}_{=a_1} \times w_z \times \underbrace{\frac{m_{z,i}}{m_{x,i}}}_{=x_i} + \underbrace{\frac{1}{w_y} \frac{M_y \sum R_y}{M_x \sum R_x}}_{=a_0} \times w_x \quad (1)$$

$$w_x = \frac{a_0}{a_1} \times w_z \quad (2)$$

where w_x is directly proportional to the mass fraction w_z of a reference material (z ; same element as the sample with the same isotopic composition as the analyte or, at least, very close to it); $m_{y,i}$, $m_{x,i}$, and $m_{z,i}$ are the masses of a spike solution (y ; same element as x with a preferably inverse isotopic composition), sample x , and reference material solution z , respectively. A number of blends (b_i ; e.g., five) containing almost the same masses of x and y , respectively, and different masses of z ($m_{z,1} < m_{z,2} < m_{z,3} < m_{z,4} < m_{z,5}$) are required for the experiment and evaluation (Figures 1 and 2). R_y , R_x , and $R_{b,i}$ denote isotope ratios measured in the respective spike, sample, and i th blend related to the isotope of major abundance (reference isotope; in the case of enriched silicon: ^{28}Si ; $R_x = I(^{30}\text{Si})/I(^{28}\text{Si})$, with the intensities I of the measured isotope signals); M_x and M_y are the molar masses of the respective analyte and spike material, which need not be known. The only requirement is the knowledge of mass fraction w_z of the element of interest in reference material z . This is given in a calibration certificate or accessible via a separate measurement (gravimetric preparation).

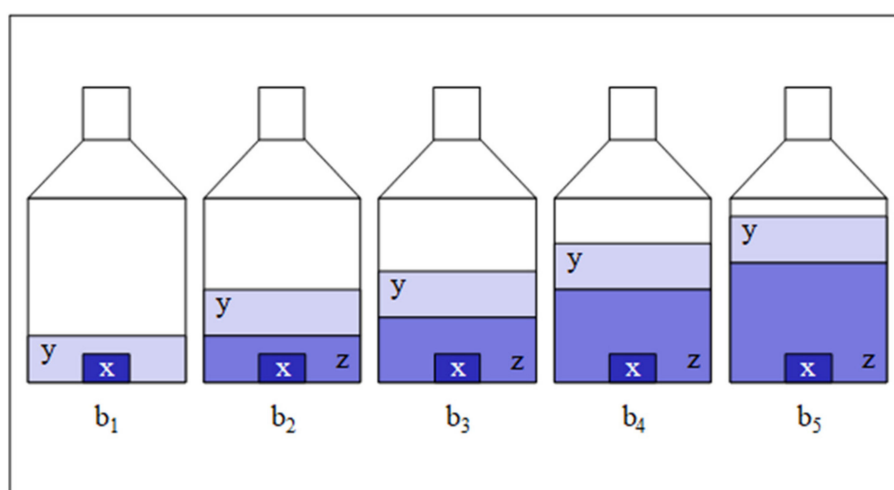


Figure 1. Schematic of a set of 5 blends (b_i) gravimetrically prepared from approximately the same masses ($m_{x,i}$) of analyte sample x , same masses ($m_{y,i}$) of spike solution y , and different masses ($m_{z,i}$) of reference solution z ($m_{z,1} < m_{z,2} < m_{z,3} < m_{z,4} < m_{z,5}$; in this work, $m_{z,1} = 0$ g).

One advantage of this new method is based on the fact that the measured intensity ratios are entered into Equation (1) directly without the need for mass bias correction. Usually, in ICP-MS, measured isotope ratios suffer from mass discrimination effects (mass bias) and have to be corrected by the use of calibration K factors. The latter are accessible via reference materials, applications of empirical relations like the exponential law, or gravimetric methods [3,5,28]. In the set of equations used in this work, however, the K factors applied to measured isotope ratios cancel out, enabling the direct use of the measured ratios and, thus, simplifying this method for convenience. The only exception is spike ratio R_y , for which no value measured in the matrix is available. Reasonable spike materials feature $R_y > 100$. In this case, mixing ratio R_y , measured in a matrix different from that of the sample matrix, with R_x and $R_{b,i}$ measured in the sample matrix, changes the result (w_x) well within its uncertainty and, therefore, is insignificant.

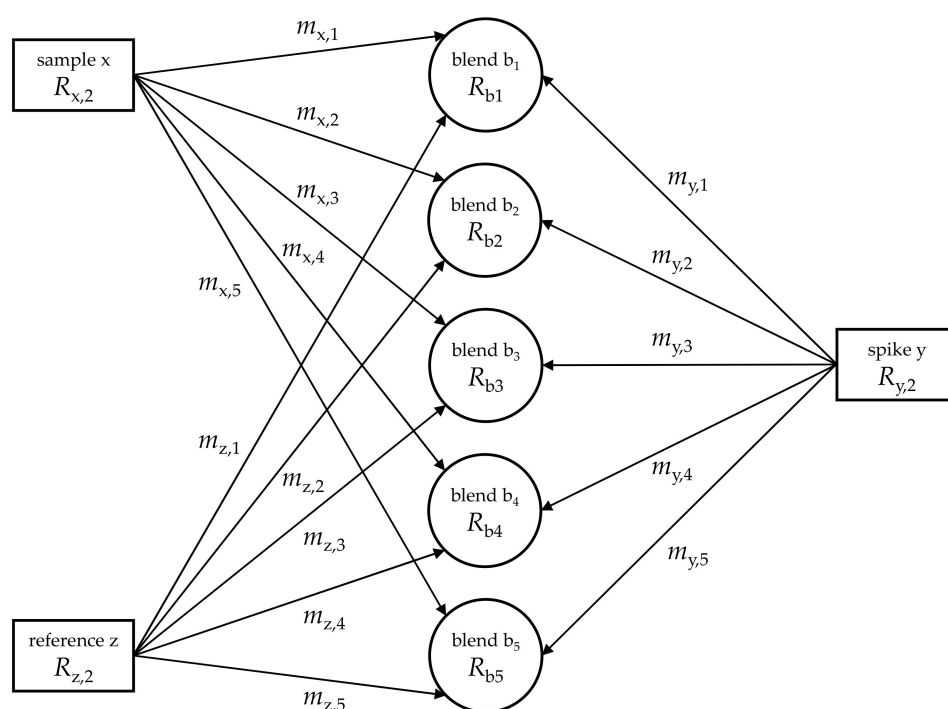


Figure 2. Relationship of the initial components (sample x, spike y, and reference z) and the respective blends (b_i), indicating (a) the abundances in which the components were blended and (b) the quantities, which have to be measured to be able to calculate analyte mass fraction w_x in the sample.

As stated above, Equation (9) in [1] has been analytically derived again in this work from scratch. This derivation is given in the Supplementary Materials. Equation (3) displays the analytical solution for mass fraction w_x in the notation used in this work. This equation is exactly the same as Equation (9) in [1]. Additionally, mass fraction w_z of the reference material used has to be known. Masses ($m_{j,i}$) of components j (j = sample x, reference z, or spike y) in blends i ($i = 1, 2, 3$) have to be known (gravimetric preparation) as well as the respective isotope ratios, namely, the R_{b3} reads:isotope ratio (monitor vs. reference isotope) in Blend 3, the $R_{z,2}$ reads:isotope ratio (monitor vs. reference isotope) in reference z, and $x_{z,1}$ reads:amount-of-substance fraction x of Isotope 1 (reference isotope) in reference material z. Note that the “reference” isotope is the isotope of the highest abundance in the sample, whereas “reference” material is a characterized material with (almost) the same isotopic composition as the sample. In the case of different isotopic compositions of x and z, the molar masses, M_x (sample) and M_z (reference), and the amount-of-substance fractions, $x_{z,1}$ and $x_{x,1}$, have to be known. When applying Equation (3), three gravimetrically prepared blends are required.

$$w_x = w_z \frac{\begin{bmatrix} m_{y1}m_{z2}m_{y3}(R_{b2} - R_{z,2})(R_{b3} - R_{b1}) \\ -m_{y1}m_{y2}m_{z3}(R_{b3} - R_{z,2})(R_{b2} - R_{b1}) \\ -m_{z1}m_{y2}m_{y3}(R_{b1} - R_{z,2})(R_{b3} - R_{b2}) \\ -m_{y1}m_{x2}m_{y3}(R_{b2} - R_{x,2})(R_{b3} - R_{b1}) \\ +m_{y1}m_{y2}m_{x3}(R_{b3} - R_{x,2})(R_{b2} - R_{b1}) \\ +m_{x1}m_{y2}m_{y3}(R_{b1} - R_{x,2})(R_{b3} - R_{b2}) \end{bmatrix}}{\begin{bmatrix} x_{z,1}M_x \\ x_{x,1}M_z \end{bmatrix}} \quad (3)$$

3. Materials and Experimental Methods

In the first test system, the sample solutions, x, consisted of aqueous TMAH ($w(\text{TMAH}) = 0.0006 \text{ g/g}$) as the matrix with traces of the analyte: silicon with natural isotopic composition. The sample solutions were taken from a study described in [29]. There, the aqueous TMAH served as a blank solution for the measurement of isotope

ratios of enriched silicon samples. Briefly, the TMAH sample solutions were prepared by dissolving highly concentrated ($w(\text{TMAH}) = 0.25 \text{ g/g}$), electronic grade (99.9999%) TMAH from Alfa Aesar™ (Thermo Fisher (Kandel) GmbH, Germany) in highly purified water (resistivity = $18 \text{ M}\Omega \text{ cm}$), the latter prepared using a commercial water purification system (Merck Millipore™, Burlington, MA, USA). All solutions were prepared and stored in precleaned labware (perfluoroalkoxy copolymer PFA) according to a protocol given in [29]. The spike solutions, y , originated from chemically pure silicon single crystals highly enriched in ^{30}Si dissolved in aqueous TMAH, whereas reference material z was prepared from well-characterized silicon crystals (material code: WASO04) with known natural-like isotopic composition [30]. All stock solutions, dilutions, and blends in this study were prepared gravimetrically, applying air buoyancy correction [31]. The silicon crystals used for the stock solutions of y (spike) and z (reference) were cleaned and etched prior to dissolution with the final mass fractions prior to blend preparation: $w_z = 4 \text{ }\mu\text{g/g}$; $w_y = 2 \text{ }\mu\text{g/g}$. The isotopic composition of y as well as the respective “true” isotope ratios, R_y , were taken from [20]. For the determination of w_x according to Equations (1) and (2), a series of five blends, b_i , was prepared, each consisting of approximately $m_{x,i} = 10 \text{ g}$, $m_{y,i} = 22.5 \text{ g}$, and stepwise increasing amounts of solution z with $m_{z,1} = 0 \text{ g}$, $m_{z,2} = 7.5 \text{ g}$, $m_{z,3} = 10 \text{ g}$, $m_{z,4} = 14.5 \text{ g}$, and $m_{z,5} = 23 \text{ g}$ (compare Figure 1).

The sample preparation for sulfur determination in the biodiesel fuel matrix is described in [23]. Briefly, an ampoule (15 mL) from the CCQM-K123 study containing commercial biodiesel fuel doped with the target analyte sulfur using dibutyl sulfide was used as the sample. Defined aliquots of reference z (NIST SRM 3154) and spike y (BAM S-34) material were added prior to the digestion. For the blend solutions, acid digestion was applied ($\text{HNO}_3/\text{H}_2\text{O}_2$) using a high-pressure asher system (Anton Paar, Graz, Austria), with $\vartheta_{\text{max}} \approx 300 \text{ }^\circ\text{C}$ and $p_{\text{max}} \approx 130 \text{ bar}$. After digestion, the solution was evaporated to dryness, redissolved with 2 mL HNO_3 (0.028 mol/L), and loaded on chromatographic columns filled with 1 mL AG 1X8 resin. The sample matrix was eluted with water, and, subsequently, the sulfur was eluted with 8 mL HNO_3 (0.25 mol/L). Samples were evaporated to dryness and redissolved in HNO_3 (0.02 kg/kg) to adjust a sulfur mass fraction of 2 mg/kg. Prior to the MC-ICP-MS measurements, sodium was added so that a final Na mass fraction of 4 mg/kg was achieved. The addition of sodium significantly increases sensitivity and prevents losses of sulfur through the membrane of the desolvating nebulizer system, presumably as sulfuric acid [32]. The complete analytical procedure was validated by applying the double IDMS calibration approach with reference material NIST SRM 2723a. The determined sulfur mass fraction was $(10.94 \pm 0.10) \text{ mg/kg}$, while the certified sulfur mass fraction was $(10.90 \pm 0.31) \text{ mg/kg}$, with $k = 2$ in both cases.

For the measurement of the third system, TRF in human serum, reference material ERM®-DA470k/IFCC “Human Serum” (Merck KGaA, Darmstadt, Germany) was applied as the reference, and Seronorm™ Immunoprotein Lyo L-1 (Invicon GmbH, Munich, Germany), which has certified values and ranges in accordance with the Guidelines of the German Medical Association (Rili-BÄK) of 2014, was used as the sample [33]. Both materials were reconstituted according to the manufacturers’ protocols and thereafter used for sample preparation [33]. The iron saturation procedure was based on the method described by del Castillo Busto et al. and C. Frank et al. except for some optimizations [15,34]. For the individual blends, a solution containing 7.5 to 22.5 mg of an iron solution (250 $\mu\text{g/g}$ in 2.5% HNO_3 , dilution from BAM A-primary-Fe-2; BAM, Berlin, Germany), 5 to 10 mg sodium carbonate (Na_2CO_3) solution (500 mmol/kg; BioUltra, Merck KGaA, Darmstadt, Germany), and 50 to 277 mg tris(hydroxymethyl)methylamine (Tris) buffer solution (12.5 mmol/kg, adjusted to $p\text{H}$ 6.4 with acetic acid; BioUltra, Merck KGaA, Darmstadt, Germany) was mixed, and 0 to 200 mg of the reconstituted ERM®-DA470k/IFCC serum and 50 mg of the reconstituted Seronorm™ Immunoprotein Lyo L-1 serum were added. The amount of iron and Na_2CO_3 solution was adjusted to the amount of added human serum, which increased from Blend 1 to Blend 4. All solutions were incubated at room temperature for 1.5 h. Then,

160 mg of the TRF spike was added. In Section 4.3, the added amounts of reference, sample, and spike solution for the different blends are given.

The TRF spike was prepared in advance from human apo-TRF purchased from Sigma-Aldrich (St Louis, MO, USA) and ^{54}Fe spike from Trace Science International Corp. (Ontario, ON, Canada), with a certified isotopic abundance of ^{54}Fe 99.86%, ^{56}Fe 0.11%, ^{57}Fe 0.01%, and ^{58}Fe 0.02%. For this, 20 mg of apo TRF was dissolved in a solution containing 0.435 g Na_2CO_3 (500 mmol/kg), 4.3 g Tris (12.5 mmol/kg), and 0.4 g ^{54}Fe (250 $\mu\text{g/g}$), adjusted to $\text{pH} = 8$ with 0.15 mmol/kg HNO_3 . After an incubation time of 3 h at room temperature, the solution was purified with a PD-10 desalting column (Cytiva GmbH, Freiburg im Breisgau, Germany), according to the manufacturer's protocol. The spike solution was freeze-dried, and the solid was used for the preparation of a TRF spike solution of 2.3 mg/g, which was used to prepare the blends.

After the gravimetric sample and blend preparation, in all blends, lipoprotein precipitation was performed by adding 5 μL of a solution containing 200 mg magnesium chloride (Merck KGaA, Darmstadt, Germany) and 100 mg dextran sulfate sodium salt (Merck KGaA, Darmstadt, Germany) in 1 mL ultrapure water, incubating for 30 min at 4 $^\circ\text{C}$ and centrifuging the samples at $12,000\times g$ for 10 min. Each supernatant was purified with PD MidiTrap columns (Cytiva GmbH, Freiburg im Breisgau, Germany), according to the manufacturer's protocol. The solutions obtained were used for the HPLC/ICP-MS analysis.

All three sample/matrix systems were investigated using ICP-MS instrumentation. The respective operation parameters of the isotope ratio measurements are summarized in Table 1. As an example, the Si in TMAH measurements were run in six sequences measuring the intensities of all three stable Si isotopes (^{28}Si , ^{29}Si , and ^{30}Si) in each blend, b_i ($i = 1 \dots 5$).

Table 1. Operation parameters of the mass spectrometric isotope ratio measurements applied for the three sample/matrix systems. (SP = spike; K = K factor).

Sample/Matrix	Silicon/TMAH	Sulfur/Biodiesel Fuel	TRF/Human Serum
Laboratory	PTB	BAM	PTB
Instrument	Thermo MC-ICP-MS Neptune	Thermo MC-ICP-MS Neptune Plus	Agilent 8900 ICP-QQQ-MS
Sample Introduction	PFA nebulizer 100 $\mu\text{L}/\text{min}$ PEEK/PFA cyclonic + Scott chamber sapphire torch + injector BN shield Ni sampler + Ni X-skimmer	Aridus II desolvating system PFA nebulizer 100 $\mu\text{L}/\text{min}$ Aridus PFA spray chamber standard torch and injector quartz shield Ni sampler + Ni H-skimmer	PFA MicroFlow nebulizer 700 μL , Scott chamber at 3 $^\circ\text{C}$ torch with 1 mm injector Pt shield Pt sampler and skimmer
Gas Flow Rates (Ar)	cooling: 16 L min^{-1} auxiliary: 0.8 L min^{-1} sample: 1.0 L min^{-1}	cooling: 16 L min^{-1} auxiliary: 0.9 L min^{-1} sample: 0.85 L min^{-1}	cooling: 15 L min^{-1} auxiliary: 0.9 L min^{-1} nebulizer gas: 0.8 L min^{-1} reaction gas (H_2): 6.1 mL min^{-1}
Machine Parameters	high resolution ($M/\Delta M = 8000$) RF power 1180 W integration time 4.194 s idle time 3 s number of blocks 6 cycles/block 3 rotating amplifiers: yes Faraday cups: L3(^{28}Si), C(^{29}Si), H3(^{30}Si)	high resolution ($M/\Delta M = 8000$) RF power 1200 W integration time 4.194 s idle time 3 s number of blocks 1 cycles/block 40 rotating amplifiers: no Faraday cups: L3(^{32}S), C(^{33}S), H3(^{34}S)	MS/MS mode RF power 1550 W Sample depth 8.0 mm x-lens configuration integration time 0.1 s m/z 53, 54, 56, 57, 58, 60

Table 1. Cont.

Sample/Matrix	Silicon/TMAH	Sulfur/Biodiesel Fuel	TRF/Human Serum
Sequence Settings	rinse time 120 s take-up time 60 s measured samples/sequence b1, b2, b3, b4, b5 (4 times each)	rinse time 30 s take-up time 80 s measured samples/sequence b1, b2, b3, b4, b5 (3 times each, separated by a block of 5 standards)	rinse time + take-up not applicable: HPLC separation measured samples/sequence b1, b2, b3, b4, blank, SP, K, blank (4 times)
Separation Settings			Agilent Bioinert 1260 HPLC system Column: MonoQ® GL 5/50 from GE Healthcare (Uppsala, Sweden) Mobile phase A: 12.5 mmol/L Tris at pH = 6.4 Mobile phase B: 12.5 mmol/L Tris + 125 mmol/L NH ₄ Ac at pH = 6.4 Flow: 0.5 mL min ⁻¹ Gradient: 0 min → 0% B, 20 min → 100% B, 27 min → 100% B Column oven 30 °C Injection volume: 10 µL MWD 254 nm, 280 nm

4. Results and Discussion

4.1. Silicon in Aqueous TMAH

The first example of the application of the new combined IDMS-Standard Addition approach is the determination of silicon with natural isotopic composition in the matrix of aqueous TMAH. As an example, the essential input and output quantities of one experimental run (Sequence 1) are given in Table 2. The corresponding linear regression curve is displayed in Figure 3.

Table 2. Determination of $w_x(\text{Si})$ in TMAH_{aq}. The relevant input and output data of the linear regression analysis are given for a single representative dataset (Sequence 1), with $w_z = 4.0069 \mu\text{g/g}$.

b_i	x TMAH _{aq}	z WASO04	y "Si30"	$R_{b,i}$	x_i	y_i	$R_{x,i}$	$R_{y,i}$
i	$m_{x,i}$	$m_{z,i}$	$m_{y,i}$	$I(^{30}\text{Si})/I(^{28}\text{Si})$	x_i	y_i	$I(^{30}\text{Si})/I(^{28}\text{Si})$	$I(^{30}\text{Si})/I(^{28}\text{Si})$
	g	g	g	V/V	g/g	g/g	V/V	mol/mol
1	10.0863	0.0000	22.8557	113.77732	0.0000	1.80	0.03353	204.19578
2	9.7836	7.8858	22.7577	1.59655	0.8060	301.51	0.03353	204.19578
3	9.6700	10.5255	22.3198	1.18847	1.0885	405.71	0.03353	204.19578
4	11.3192	15.3000	22.4864	0.83531	1.3517	503.86	0.03353	204.19578
5	10.0440	22.4061	22.7651	0.61405	2.2308	794.84	0.03353	204.19578
					a_1	a_0	w_x	
					(g/g)/(g/g)	(g/g)	µg/g	
					356.10062	11.474	0.13	

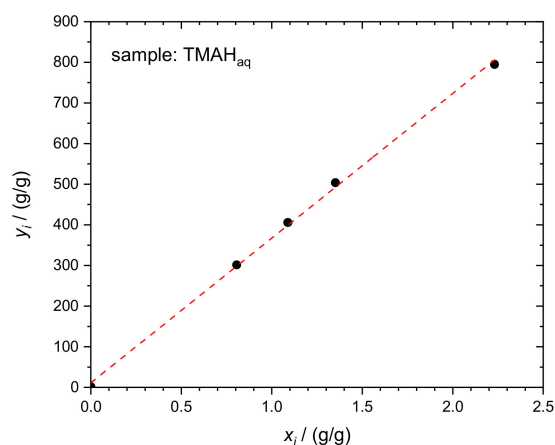


Figure 3. Linear regression evaluation of the Si mass fraction in TMAH_{aq} according to Equation (2). Dataset in Table 2.

As a direct consequence of linear regression analysis, mass fraction w_x of the analyte in the respective sample x (TMAH_{aq}) was derived according to Equation (2). A set of 5 blends is sufficient to obtain proper regression statistics.

The individual results of $w_x(\text{Si})$, obtained from six sequences (runs), are displayed in Figure 4 (left) and Table 3. They yield an average of $w_x(\text{Si}) = 0.081(73) \mu\text{g/g}$. As proof of consistency of the individual measurement results, the concept of degrees of equivalence (d_i) was applied. The individual d_i of the respective measurements are plotted in Figure 4 (right). All data encompass the zero line with their associated expanded uncertainties, which is an approval of the complete consistency of the data.

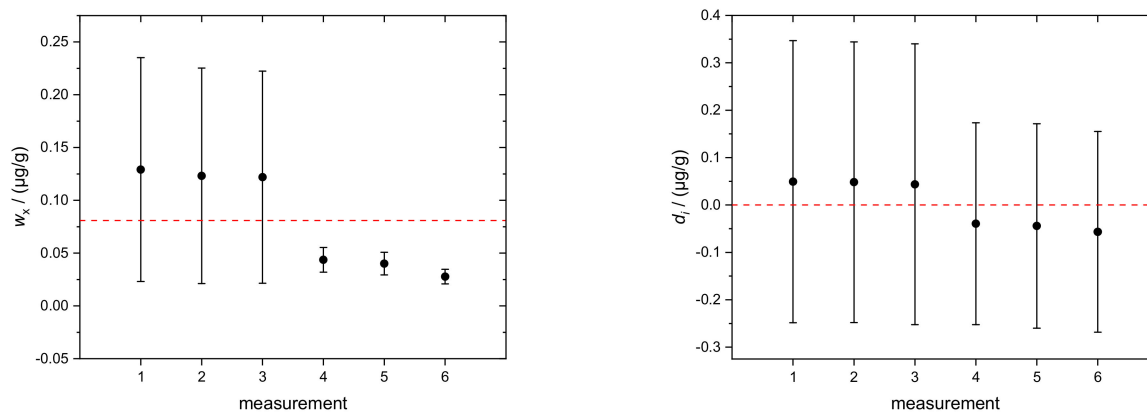


Figure 4. (Left): Mass fraction w_x (Si) in TMAH_{aq}. Error bars denote combined uncertainties ($k = 1$). The red dashed line indicates the average value. (Right): Degrees of equivalence d_i of the respective measurement results. Error bars indicate expanded uncertainties ($k = 2$) associated with d_i . All single results are consistent with the average value since the respective uncertainties encompass the red dashed zero line.

Table 3. Mass fractions w_x (Si) of silicon in TMAH_{aq} and the associated uncertainties ($k = 1$).

Run	w_x (Si) $\mu\text{g/g}$	$u(w_x$ (Si)) $\mu\text{g/g}$
1	0.13	0.11
2	0.12	0.10
3	0.12	0.10
4	0.044	0.012
5	0.040	0.011
6	0.028	0.007
average	0.081	0.073

4.2. Sulfur in Biodiesel Fuel (BDF)

The determination of sulfur in BDF, according to Equations (1) and (2), was performed in a similar way using the five blends, b_i . The essential input and output quantities of one experimental run are given in Table 4. The corresponding linear regression curve of the respective dataset is displayed in Figure 5.

Table 4. Determination of w_x in BDF. The relevant input and output data of linear regression analysis are given for a single representative dataset (1st run), with $w_z = 16.145 \mu\text{g/g}$. A procedural blank was subtracted from the result. The procedural blank was determined during an external measurement, as described in [23].

b_i	x BDF	z NIST SRM 3154	y BAM S-34	$R_{b,i}$	x_i	y_i	$R_{x,i}$	$R_{y,i}$
i	$m_{x,i}$ g	$m_{z,i}$ g	$m_{y,i}$ g	$I(^{32}\text{S})/I(^{34}\text{S})$ V/V	x_i g/g	y_i g/g	$I(^{32}\text{S})/I(^{34}\text{S})$ V/V	$I(^{32}\text{S})/I(^{34}\text{S})$ mol/mol
1	0.23748	0.00000	0.09670	0.20030	0.00000	0.00387	21.16643	0.00099
2	0.24149	0.10125	0.09843	0.36731	0.41927	0.00718	21.16643	0.00099
3	0.23819	0.20828	0.10899	0.48453	0.87443	0.01070	21.16643	0.00099
4	0.25126	0.30375	0.10631	0.64994	1.20891	0.01339	21.16643	0.00099
5	0.24311	0.40679	0.09966	0.83887	1.67328	0.01690	21.16643	0.00099
					a_1 (g/g)/(g/g) 0.007797	a_0 g/g 0.003894	w_x $\mu\text{g/g}$ 8.063	$w_{x,\text{corr}}$ $\mu\text{g/g}$ 7.36

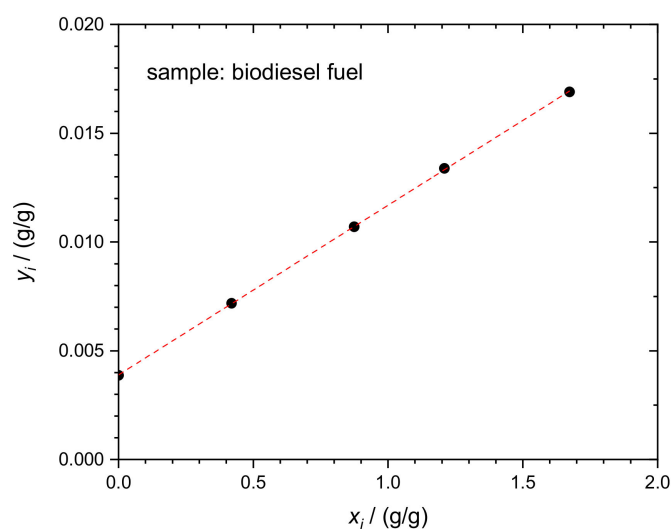


Figure 5. Linear regression of the S mass fraction in biodiesel fuel according to Equation (2). Dataset in Table 4.

Linear regression analysis yields the mass fraction (w_x) of the analyte (sulfur) in the respective sample x (BDF), according to Equation (2).

The determination of sulfur in BDF, according to Equations (1) and (2), yields an average mass fraction of $w_{x,\text{corr}}(\text{S}) = 7.36(11) \mu\text{g/g}$. The individual results of $w_{x,\text{corr}}(\text{S})$, measured in three runs, are shown in Figure 6 (left) and Table 5. Additionally, as proof of consistency, the concept of degrees of equivalence, d_i , was applied. The d_i of the respective measurements are plotted in Figure 6 (right). The single results encompass the zero line with their associated expanded uncertainties, which proves the complete consistency of the data. In the case of sulfur determination in this work, the respective sample was taken from a stock solution already used in the interlaboratory comparison CCQM-K123, as described in [23]. There, the sulfur determination was carried out using double IDMS, yielding $w_x = 7.394(46) \mu\text{g/g}$. The corresponding result is shown in Figure 6 (left, red solid

line) for comparison. Both the IDMS and the new combined IDMS/standard addition approach show excellent agreement within the limits of uncertainty.

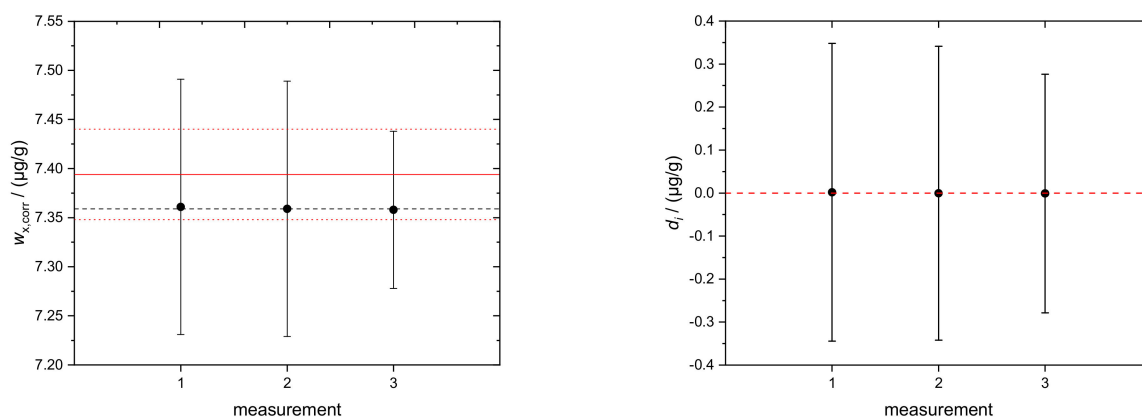


Figure 6. (Left) Black circles: corrected mass fractions $w_{x,corr}(S)$ of sulfur in BDF and the associated uncertainties ($k = 1$); black dashed line: average of three single runs. Data from the new combined IDMS/standard addition approach (this work). Red solid line: average value of $w(S)$ from BAM, applying IDMS and using the same solution [23]. Upper and lower associated uncertainties: red dotted lines. (Right) Degrees of equivalence (d_i) of the respective measurement results. Error bars indicate expanded uncertainties ($k = 2$) associated with d_i . All single results are consistent with the average value since the respective uncertainties encompass the red dashed zero line.

Table 5. Blank corrected mass fractions $w_{x,corr}(S)$ of sulfur in biodiesel fuel (uncertainties with $k = 1$).

Run	$w_{x,corr}(S)$ $\mu\text{g/g}$	$u(w_{x,corr}(S))$ $\mu\text{g/g}$
1	7.36	0.13
2	7.36	0.13
3	7.358	0.079
average	7.36	0.11

4.3. Transferrin in Human Serum

The determination of TRF in human serum, according to Equations (1) and (2), was performed in a similar way using four blends, b_i . The essential input and output quantities of one experimental run are given in Table 6. The corresponding linear regression curve of the respective dataset is displayed in Figure 7.

Table 6. Determination of $w_x(\text{TRF})$ in human serum. The relevant input and output data of linear regression analysis are given for a single representative dataset (M21-3), with $w_z = 2.296 \text{ mg/g}$.

b_i	x Seronorm TM Immuno- Protein Lyo L-1	z ERM [®] - DA470k/IFCC	y In-House Prepared TRF Spike	$R_{b,i}$	x_i	y_i	$R_{x,i}$	$R_{y,i}$
i	$m_{x,i}$ g	$m_{z,i}$ g	$m_{y,i}$ g	$R(^{54}\text{Fe}/^{56}\text{Fe})$ mol/mol	x_i g/g	y_i g/g	$R(^{54}\text{Fe}/^{56}\text{Fe})$ mol/mol	$R(^{54}\text{Fe}/^{56}\text{Fe})$ mol/mol
1				3.01104	0.00000	262.4		
2				2.99958	0.00000	263.4		
3	0.04848	0.00000	0.15104	2.99154	0.00000	264.1		
4				2.98887	0.00000	264.4		
5				1.31282	1.18510	614.6		
6	0.04923	0.05834	0.15123	1.32126	1.18510	610.5		
7				1.31656	1.18510	612.8		
8				1.31365	1.18510	614.2	0.063703	251.22
9				1.15279	1.51220	697.3		
10	0.04910	0.07424	0.14909	1.15426	1.51220	696.3		
11				1.15733	1.51220	694.4		
12				1.14892	1.51220	699.8		
13				0.74959	2.95788	1109.1		
14	0.04923	0.14561	0.14951	0.74722	2.95788	1113.0		
15				0.74440	2.95788	1117.6		
16				0.74494	2.95788	1116.7		
					a_1 (g/g)/(g/g) 287.078	a_0 g/g 266.04	w_x mg/g 2.128	

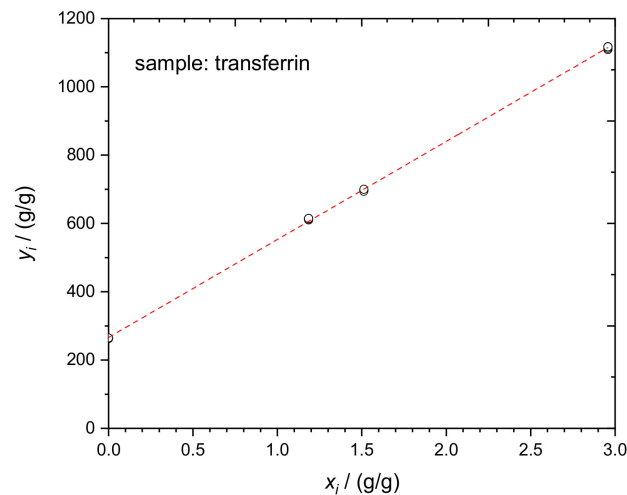


Figure 7. Linear regression evaluation of the TRF mass fraction in human serum according to Equation (2). Dataset in Table 6.

The linear regression analysis yields mass fraction w_x of the analyte TRF in the respective sample x (human serum) according to Equation (2).

The individual results of $w_x(\text{TRF})$, obtained from two independent measurement sequences, are displayed in Figure 8 (left). Both values, $w_x(\text{TRF}) = 2.128(45) \text{ mg/g}$ and $w_x(\text{TRF}) = 2.340(76) \text{ mg/g}$ for $k = 1$, are consistent with the certified target value and its allowed range [33]. As proof of consistency of the individual measurement results, the concept of degrees of equivalence, d_i , was applied. The individual d_i of the respective

measurements are plotted in Figure 8 (right). All data encompass the zero line with their associated expanded uncertainties.

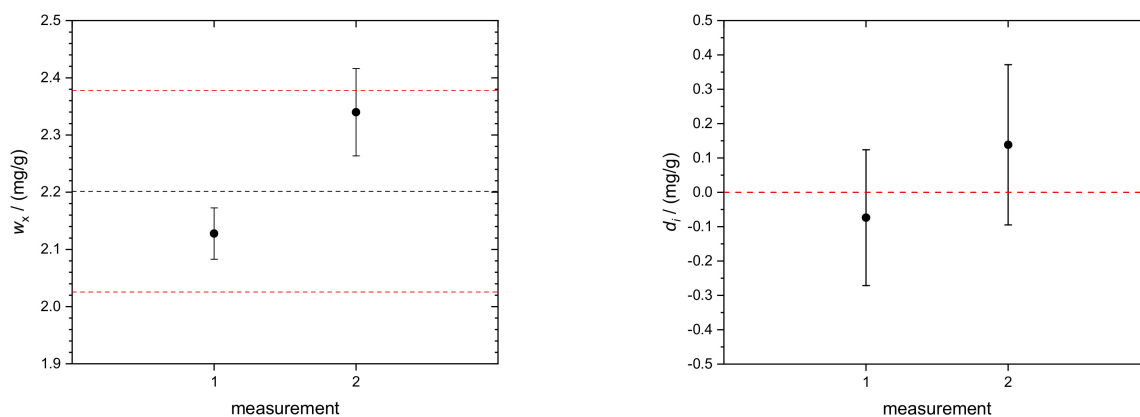


Figure 8. (Left) Black circles: mass fractions w_x (TRF) of transferrin in human serum and the associated uncertainties ($k = 1$); dashed black line: certified value of the Seronorm sample (for instrument, see Beckmann, AU). Data from the new combined IDMS/standard addition approach (this work). Red dashed lines: allowed target range according to Rili-BÄK [33]. (Right) Degrees of equivalence, d_i , of the respective measurement results. Error bars indicate expanded uncertainties ($k = 2$) associated with d_i . All single results are consistent with the certified value since the respective uncertainties encompass the red dashed zero line.

4.4. Comparison of Linear Regression (This Work) and Analytical Solution (Pagliano and Meija)

The comparison of the two approaches, combining IDMS and standard addition presented in this work and in [1], was performed using the dataset of silicon determination in the TMAH matrix (five available blends: b_1, b_2, b_3, b_4, b_5). Figure 9 shows the results of w_x (Si), determined in six sequences (runs), using five blends (this work) and three blends (b_2, b_3, b_4 ; approach of [1] using Equation (3)). At first glance, the associated uncertainties of the results determined in this work (black circles) are significantly smaller than the respective uncertainties of the results determined using the approach of [1] (red circles).

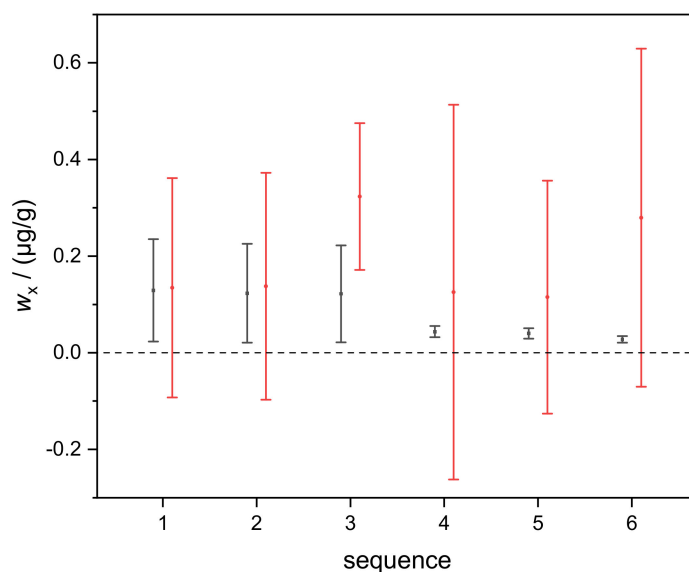


Figure 9. Black circles (error bars): mass fractions w_x (Si) and the associated uncertainties ($k = 1$) of six runs determined using the approach of this work. Red circles (error bars): results of the same input data (blends b_2, b_3, b_4 only) using the approach of [1].

Prior to this comparison, both approaches (this work and that of [1]) were checked for consistency using simulated ideal data: both methods yielded exactly the same result. However, when using real experimental data (e.g., the results of the five blends for silicon determination), the results of the approach of [1] depend strongly on the input data (the three blends that were used for analysis). Table 7 shows the different numerical results between the two approaches of a representative run. In the case of this work, all five blends were used (left column), whereas, for the approach of [1], triple blends b_1, b_2, b_3 or b_2, b_3, b_4 , or b_3, b_4, b_5 each yielded extremely different results. The analytical approach (Equation (9) of [1], which is equal to Equation (3) in this publication) is based on a set of three blends only, without the need for the measurement of R_y , which appears initially as a benefit. However, the analysis shows that the three-blend application leads, in some cases, to unrealistic (not accurate) results (results of triples b_1, b_2, b_3 and b_3, b_4, b_5). Equation (9) in [1] reacts extremely sensitively towards that kind of input data. In our case, the three inner blends yield the most reasonable (“accurate”) result, e.g., the first blend, b_1 , includes no reference material z ($m_z = 0$ g). This suggests a rather unstable and sensitive applicability of Equation (9) of [1] compared to the linear regression approach of this work. Moreover, for practical applications, Equations (1) and (2) are much simpler and user-friendly, and the respective evaluations and measurements are much easier than in the case of the approach used in [1], although the latter is correct from the mathematical point of view.

Table 7. A representative result of mass fractions $w_x(\text{Si})$ determined using the approach of this work (using all five blends) and the approach of [1] (using the three triple blends: b_1, b_2, b_3 ; b_2, b_3, b_4 ; and b_3, b_4, b_5). Only the combination of b_2, b_3, b_4 (the inner blends) yielded a reasonable numerical result.

This Work	Approach of [1]
blends	blends
b_1, b_2, b_3, b_4, b_5	b_1, b_2, b_3
$w_x(\text{Si})$	$w_x(\text{Si})$
$\mu\text{g/g}$	$\mu\text{g/g}$
0.1292	−0.5004

For the comparison of the uncertainties associated with w_x , which is the second-most important outcome of the respective approach, we chose the results of the blend triple b_2, b_3, b_4 of the six sequences. As can be seen in Figure 9, the uncertainties obtained using the approach of this work are significantly smaller than those obtained using Equation (9) from [1].

An uncertainty analysis (Table 8) of Equation (3) (same as Equation (9) in [1]) according to GUM [9], using GUM Workbench Pro™ software (version 2.4.1 392, Metrodata GmbH, Germany), shows that the absolute values of the sensitivity coefficients of the main uncertainty contributions, R_{b_2} , R_{b_1} , and R_{b_3} , range in the 10^1 - 10^2 region. This seems rather high and is mainly responsible for the elevated uncertainty compared to the smaller uncertainty obtained using linear regression in the approach of this work.

Table 8. A representative uncertainty budget of mass fraction $w_x(\text{Si})$ determined using the approach of [1] (using the blends b_2, b_3, b_4). The main contributions originate from $R_{b2}, R_{b1},$ and R_{b3} , with the largest absolute values of respective sensitivity coefficients.

Quantity	Unit	Best Estimate (Value)	Standard Uncertainty	Sensitivity Coefficient	Index
X_i	$[X_i]$	x_i	$u(x_i)$	c_i	
w_z	$\mu\text{g/g}$	4.00694	6.01×10^{-3}	0.031	0.0%
m_{y1}	g	22.49660	1.00×10^{-3}	2.2	0.0%
m_{z2}	g	10.31270	1.00×10^{-3}	11	0.0%
m_{y3}	g	22.26850	1.00×10^{-3}	2.8	0.0%
R_{b2}	V/V	1.23933	3.62×10^{-3}	92	74.4%
R_{z2}	V/V	0.033527	335×10^{-6}		
R_{b3}	V/V	0.88472	1.50×10^{-3}	−73	8.0%
R_{b1}	V/V	1.68846	5.46×10^{-3}	−30	17.5%
m_{y2}	g	22.43470	1.00×10^{-3}	−5.0	0.0%
m_{z3}	g	14.57170	1.00×10^{-3}	−4.2	0.0%
m_{z1}	g	7.46840	1.00×10^{-3}	−6.4	0.0%
m_{x2}	g	9.22730	1.00×10^{-3}	0.33	0.0%
R_{x2}	V/V	0.033527	335×10^{-6}	10	0.0%
m_{x3}	g	9.79000	1.00×10^{-3}	−0.13	0.0%
m_{x1}	g	9.46490	1.00×10^{-3}	−0.20	0.0%
w_x	g	0.125	0.388		

5. Conclusions

The new method presented—a combination of IDMS and standard addition—for absolute elemental quantification is recommended for sample analytes with two or more isotopes, especially when appearing in traces (ng/g) and/or in a complex matrix that is difficult or (almost) impossible to separate. The three different examples of this study demonstrate the convenient applicability of this approach successfully. Moreover, the second example (determination of sulfur in biodiesel fuel) acts as a validation system since the same sample material has been measured previously and independently using a double IDMS method in the context of an international key comparison (CCQM-K123), yielding excellently matching results within the limits of uncertainty. Mainly, since the blends are measured, all the “measurement” solutions are matrix-matched; thus, potential matrix effects are also canceled out. The comparison of the new method (linear regression) with the analytical approach reported in [1] shows the formal (mathematical) agreement of both methods. However, the use of experimental data suggests a more robust behavior of the approach presented in this work, yielding more “accurate” results with significantly lower associated uncertainties (see calculation examples in the Supplement). Moreover, the new approach is more practical in daily laboratory work due to a simpler evaluation algorithm that might be less fault-prone.

Supplementary Materials: The following are available online, Supplement_a_ID_STDAdd.docx: with Table S1: List of quantities and symbols in the general final Equation (3) and Supplement-b-StdAdd-IDMS.xlsm.

Author Contributions: Conceptualization, A.P. and O.R.; methodology, O.R.; validation, A.P. and J.V.; formal analysis, C.B., A.P., O.R., A.S., P.P., and J.V.; investigation, C.B., A.P., O.R., and J.V.; data curation, C.B., A.P., O.R., and J.V.; writing—original draft preparation, A.P.; writing—review and editing, C.B., A.P., O.R., and J.V. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Data Availability Statement: The data that support the findings of this study are available from the authors upon request.

Acknowledgments: The authors gratefully acknowledge animated discussions with Detlef Schiel (PTB). Special thanks to Dorit Becker and Maren Koenig for their solid support with the sulfur measurements.

Conflicts of Interest: The authors declare no conflict of interest.

Appendix A

Derivation of Combined IDMS and Standard Addition Method

Isotope ratios ($R_{b,i}$) in the blends (i) containing the analyte can be expressed as the ratio of the sums of the amount-of-substances (n) of the respective components of x (sample), y (spike), and z (reference). In our notation, Subscript 1 denotes the reference and Subscript 2 the monitor (spike) isotope.

$$R_{b,i} = \frac{n_{x2,i} + n_{z2,i} + n_{y2,i}}{n_{x1,i} + n_{z1,i} + n_{y1,i}} \quad (\text{A1})$$

$$R_{b,i} = \frac{x_{x2}n_{x,i} + x_{z2}n_{z,i} + x_{y2}n_{y,i}}{x_{x1}n_{x,i} + x_{z1}n_{z,i} + x_{y1}n_{y,i}} \quad (\text{A2})$$

$$R_{b,i} = \frac{x_{x2} \frac{m_{x,i}w_x}{M_x} + x_{z2} \frac{m_{z,i}w_z}{M_z} + x_{y2} \frac{m_{y,i}w_y}{M_y}}{x_{x1} \frac{m_{x,i}w_x}{M_x} + x_{z1} \frac{m_{z,i}w_z}{M_z} + x_{y1} \frac{m_{y,i}w_y}{M_y}} \quad (\text{A3})$$

In case of equal isotopic patterns in the sample and reference (the equation, valid in the case of different isotopic patterns, including its derivation, is presented in the Supplement):

$$x_{x1} = x_{z1}, x_{x2} = x_{z2}, M_x = M_z \quad (\text{A4})$$

Therefore, $R_{b,i}$ can be rewritten accordingly

$$R_{b,i} = \frac{\frac{x_{x2}}{M_x} (m_{x,i}w_x + m_{z,i}w_z) + \frac{x_{y2}}{M_y} m_{y,i}w_y}{\frac{x_{x1}}{M_x} (m_{x,i}w_x + m_{z,i}w_z) + \frac{x_{y1}}{M_y} m_{y,i}w_y} \quad (\text{A5})$$

After rearranging and expanding

$$R_{b,i} \left[\frac{x_{x1}}{M_x} (m_{x,i}w_x + m_{z,i}w_z) + \frac{x_{y1}}{M_y} m_{y,i}w_y \right] = \frac{x_{x2}}{M_x} (m_{x,i}w_x + m_{z,i}w_z) + \frac{x_{y2}}{M_y} m_{y,i}w_y \quad (\text{A6})$$

$$R_{b,i} \frac{x_{x1}}{M_x} (m_{x,i}w_x + m_{z,i}w_z) + R_{b,i} \frac{x_{y1}}{M_y} m_{y,i}w_y = \frac{x_{x2}}{M_x} (m_{x,i}w_x + m_{z,i}w_z) + \frac{x_{y2}}{M_y} m_{y,i}w_y \quad (\text{A7})$$

$$(m_{x,i}w_x + m_{z,i}w_z) \frac{1}{M_x} (R_{b,i}x_{x1} - x_{x2}) = m_{y,i}w_y \frac{1}{M_y} (x_{y2} - R_{b,i}x_{y1}) \quad (\text{A8})$$

$$m_{y,i}w_y \frac{1}{M_y} (x_{y2} - R_{b,i}x_{y1}) = (m_{x,i}w_x + m_{z,i}w_z) \frac{1}{M_x} (R_{b,i}x_{x1} - x_{x2}) \quad (\text{A9})$$

$$m_{y,i} \frac{x_{y2} - R_{b,i}x_{y1}}{R_{b,i}x_{x1} - x_{x2}} = \frac{1}{w_y} \frac{M_y}{M_x} (m_{x,i}w_x + m_{z,i}w_z) \quad (\text{A10})$$

$$m_{y,i} \frac{x_{y2} - R_{b,i}x_{y1}}{R_{b,i}x_{x1} - x_{x2}} = \frac{1}{w_y} \frac{M_y}{M_x} m_{x,i}w_x + \frac{1}{w_y} \frac{M_y}{M_x} m_{z,i}w_z \quad (\text{A11})$$

$$\frac{m_{y,i}}{m_{x,i}} \frac{x_{y2} - R_{b,i}x_{y1}}{R_{b,i}x_{x1} - x_{x2}} = \frac{1}{w_y} \frac{M_y}{M_x} \frac{m_{z,i}w_z}{m_{x,i}} + \frac{1}{w_y} \frac{M_y}{M_x} w_x \quad (\text{A12})$$

$$\frac{m_{y,i}}{m_{x,i}} \frac{\frac{R_{y2}}{\sum R_y} - R_{b,i} \frac{R_{y1}}{\sum R_y}}{\frac{R_{x1}}{\sum R_x} - \frac{R_{x2}}{\sum R_x}} = \frac{1}{w_y} \frac{M_y}{M_x} \frac{m_{z,i}w_z}{m_{x,i}} + \frac{1}{w_y} \frac{M_y}{M_x} w_x \text{ with } R_{y1} = R_{x1} = 1 \quad (\text{A13})$$

$$\frac{m_{y,i}}{m_{x,i}} \frac{\frac{1}{\sum R_y} R_{y2} - R_{b,i} \times 1}{\frac{1}{\sum R_x} R_{b,i} \times 1 - R_{x2}} = \frac{1}{w_y} \frac{M_y}{M_x} \frac{m_{z,i} w_z}{m_{x,i}} + \frac{1}{w_y} \frac{M_y}{M_x} w_x \quad (\text{A14})$$

$$\frac{m_{y,i}}{m_{x,i}} \frac{\sum R_x}{\sum R_y} \frac{R_{y2} - R_{b,i} \times 1}{R_{b,i} \times 1 - R_{x2}} = \frac{1}{w_y} \frac{M_y}{M_x} \frac{m_{z,i} w_z}{m_{x,i}} + \frac{1}{w_y} \frac{M_y}{M_x} w_x \quad (\text{A15})$$

Equation (1) already represents the linear equation required to describe the regression line.

$$\underbrace{\frac{m_{y,i}}{m_{x,i}} \times \frac{R_y - R_{b,i}}{R_{b,i} - R_x}}_{=y_i} = \underbrace{\frac{1}{w_y} \frac{M_y}{M_x} \frac{\sum R_y}{\sum R_x}}_{=a_1} \times w_z \times \underbrace{\frac{m_{z,i}}{m_{x,i}}}_{=x_i} + \underbrace{\frac{1}{w_y} \frac{M_y}{M_x} \frac{\sum R_y}{\sum R_x} \times w_x}_{=a_0}$$

The y -axis intercept a_0 and slope a_1 yield the mass fraction (w_x) of the analyte element in the sample

$$w_x = \frac{a_0}{a_1} \times w_z$$

References

- Pagliano, E.; Meija, J. Reducing the matrix effects in chemical analysis: Fusion of isotope dilution and standard addition methods. *Metrologia* **2016**, *53*, 829–834. [[CrossRef](#)]
- Sargent, M.; Goenaga-Infante, H.; Inagaki, K.; Ma, L.; Meija, J.; Pramann, A.; Rienitz, O.; Sturgeon, R.; Vogl, J.; Wang, J.; et al. The role of ICP-MS in inorganic chemical metrology. *Metrologia* **2019**, *56*, 1–32. [[CrossRef](#)]
- Vanhaecke, F.; Degryse, P. *MC-ICP-MS in Isotopic Analysis: Fundamentals and Applications Using ICP-MS*; Wiley-VCH: Weinheim, Germany, 2012.
- Yang, L. Accurate and precise determination of isotopic ratios by MC-ICP-MS: A review. *Mass Spectrom. Rev.* **2009**, *28*, 990–1011. [[CrossRef](#)] [[PubMed](#)]
- Yang, L.; Tong, S.; Zhou, Z.; Hu, Z.; Mester, Z.; Meija, J. A critical review on isotopic fractionation correction methods for accurate isotope amount ratio measurements by MC-ICP-MS. *J. Anal. At. Spectrom.* **2018**, *33*, 1849–1861. [[CrossRef](#)]
- Andr n, H.; Rodushkin, I.; Stenberg, A.; Malinovski, D.; Baxter, D.C. Sources of mass bias and isotope ratio variation in multi-collector ICP-MS: Optimization of instrumental parameters based on experimental observations. *J. Anal. At. Spectrom.* **2004**, *19*, 1217–1224. [[CrossRef](#)]
- Pramann, A.; Rienitz, O.; Schiel, D.; G ttler, B.; Valkiers, S. Novel concept for the mass spectrometric determination of absolute isotopic abundances with improved measurement uncertainty: Part 3—Molar mass of silicon highly enriched in ^{28}Si . *Int. J. Mass Spectrom.* **2011**, *305*, 58–68. [[CrossRef](#)]
- Vogl, J. Calibration strategies and quality assurance. In *Inductively Coupled Plasma Mass Spectrometry Handbook*; Nelms, S.M., Ed.; Blackwell Publishing Ltd: Oxford, UK, 2005; pp. 147–181.
- Working Group 1 of the Joint Committee for Guides in Metrology (JCGM/WG 1). *JCGM 100:2008. Evaluation of Measurement Data—Guide to the Expression of Uncertainty in Measurement*; JCGM: Sevres, France, 2008.
- Heumann, K.G. Isotope-dilution mass spectrometry of inorganic and organic substances. *Fresenius Z. Anal. Chem.* **1986**, *325*, 661–666. [[CrossRef](#)]
- De Bi vre, P. Isotope dilution mass spectrometry: What can it contribute to accuracy in trace analysis? *Fresenius Z. Anal. Chem.* **1990**, *327*, 766–771. [[CrossRef](#)]
- Heumann, K.G. Isotope dilution mass spectrometry (IDMS) of the elements. *Mass Spectrom. Rev.* **1992**, *11*, 41–67. [[CrossRef](#)]
- Sargent, M.; Harrington, C.; Harte, R. *Guidelines for Achieving High Accuracy in Isotope Dilution Mass Spectrometry (IDMS)*; LGC Ltd: Cambridge, MA, USA, 2002.
- Vogl, J. Measurement uncertainty in single, double, and triple isotope dilution mass spectrometry. *Rapid Comm. Mass Spectrom.* **2012**, *26*, 275–281. [[CrossRef](#)]
- Frank, C.; Rienitz, O.; Swart, C.; Schiel, D. Improving species-specific IDMS: The advantages of triple IDMS. *Anal. Bioanal. Chem.* **2013**, *405*, 1913–1919. [[CrossRef](#)]
- Hauswaldt, A.-L.; Rienitz, O.; J hrling, R.; Fischer, N.; Schiel, D.; Labarraque, G.; Magnusson, B. Uncertainty of standard addition experiments: A novel approach to include the uncertainty associated with the standard in the model equation. *Accred. Qual. Assur.* **2012**, *17*, 129–138. [[CrossRef](#)]
- Rienitz, O.; Hauswaldt, A.-L.; J hrling, R. Standard addition challenge. *Anal. Bioanal. Chem.* **2012**, *403*, 2461–2462. [[CrossRef](#)] [[PubMed](#)]
- Rienitz, O.; Hauswaldt, A.-L.; J hrling, R. Solution to standard addition challenge. *Anal. Bioanal. Chem.* **2012**, *404*, 2117–2118. [[CrossRef](#)] [[PubMed](#)]

19. Fujii, K.; Bettin, H.; Becker, P.; Massa, E.; Rienitz, O.; Pramann, A.; Nicolaus, A.; Kuramoto, N.; Busch, I.; Borys, M. Realization of the kilogram by the XRCD method. *Metrologia* **2016**, *53*, A19–A45. [[CrossRef](#)]
20. Pramann, A.; Lee, K.S.; Noordmann, J.; Rienitz, O. Probing the homogeneity of the isotopic composition and molar mass of the ‘Avogadro’-crystal. *Metrologia* **2015**, *52*, 800–810. [[CrossRef](#)]
21. Bartl, G.; Becker, P.; Beckhoff, B.; Bettin, H.; Beyer, E.; Borys, M.; Busch, I.; Cibik, L.; D’Agostino, G.; Darlatt, E.; et al. A new ²⁸Si single crystal: Counting the atoms for the new kilogram definition. *Metrologia* **2017**, *54*, 693–715. [[CrossRef](#)]
22. Güttler, B.; Rienitz, O.; Pramann, A. The avogadro constant for the definition and realization of the mole. *Ann. Phys.* **2018**, *1800292*. [[CrossRef](#)]
23. Kuroiwa, T.; Zhu, Y.; Inagaki, K.; Long, S.E.; Christopher, S.J.; Puelles, M.; Borinsky, M.; Hatamleh, N.; Murby, J.; Merrick, J.; et al. Report of the CCQM-K123: Trace elements in biodiesel fuel. *Metrologia* **2017**, *54*, 08008. [[CrossRef](#)]
24. Jou, Y.J.; Lin, C.D.; Lai, C.H.; Chen, C.H.; Kao, J.Y.; Chen, S.Y.; Tsai, M.H.; Huang, S.H.; Lin, C.W. Proteomic identification of salivary transferrin as a biomarker for early detection of oral cancer. *Anal. Chim. Acta* **2010**, *681*, 41–48. [[CrossRef](#)]
25. Deutsches Ärzteblatt. Neufassung der Richtlinie der Bundesärztekammer zur Qualitätssicherung laboratoriumsmedizinischer Untersuchungen—Rili-BÄK. *Dtsch. Arztebl.* **2019**, *116*, A-2422 / B-1990 / C-1930.
26. Feng, L.; Zhang, D.; Wang, J.; Li, H. Simultaneous quantification of proteins in human serum via sulfur and iron using HPLC coupled to post-column isotope dilution mass spectrometry. *Anal. Methods* **2014**, *6*, 7655–7662. [[CrossRef](#)]
27. Larios, R.; Del Castillo Busto, M.E.; Garcia-Sar, D.; Ward-Deitrich, C.; Goenaga-Infante, H. Accurate quantification of carboplatin adducts with serum proteins by monolithic chromatography coupled to ICPMS with isotope dilution analysis. *J. Anal. At. Spectrom.* **2019**, *34*, 729–740. [[CrossRef](#)]
28. Mana, G.; Rienitz, O. The calibration of Si-isotope ratio measurements. *Int. J. Mass Spectrom.* **2010**, *291*, 55–60. [[CrossRef](#)]
29. Rienitz, O.; Pramann, A.; Vogl, J.; Lee, K.-S.; Yim, Y.-H.; Malinovskiy, D.; Hill, S.; Dunn, P.; Goenaga-Infante, H.; Ren, T.; et al. The comparability of the determination of the molar mass of silicon highly enriched in ²⁸Si: Results of the CCQM-P160 interlaboratory comparison and additional external measurements. *Metrologia* **2020**, *57*, 1–13. [[CrossRef](#)]
30. D’Agostino, G.; Di Luzio, M.; Mana, G.; Oddone, M.; Pramann, A.; Prata, M. ³⁰Si mole fraction of a silicon material highly enriched in ²⁸Si determined by instrumental neutron activation analysis. *Anal. Chem.* **2015**, *87*, 5716–5722. [[CrossRef](#)]
31. Kaltenbach, A.; Noordmann, J.; Görlitz, V.; Pape, C.; Richter, S.; Kipphardt, H.; Kopp, G.; Jährling, R.; Rienitz, O.; Güttler, B. Gravimetric preparation and characterization of primary reference solutions of molybdenum and rhodium. *Anal. Bioanal. Chem.* **2015**, *407*, 3093–3102. [[CrossRef](#)]
32. Paris, G.; Sessions, A.L.; Subhas, A.V.; Adkins, J.F. MC-ICP-MS measurement of $\delta^{34}\text{S}$ and $\Delta^{33}\text{S}$ in small amounts of dissolved sulfate. *Chem. Geol.* **2013**, *345*, 50–61. [[CrossRef](#)]
33. Deutsches Ärzteblatt. Neufassung der Richtlinie der Bundesärztekammer zur Qualitätssicherung laboratoriumsmedizinischer Untersuchungen—Rili-BÄK. *Dtsch. Arztebl.* **2014**, *111*, A-1583/B-1363/C-1295.
34. Del Castillo Busto, M.E.; Montes-Bayón, M.; Sans-Mendel, A. Accurate determination of human serum transferrin isoforms: Exploring metal-specific isotope dilution analysis as a quantitative proteomic tool. *Anal. Chem.* **2006**, *78*, 8218–8226. [[CrossRef](#)]